

UNIVERSITY OF WASHINGTON
DEPARTMENT OF OCEANOGRAPHY
Seattle, 5, Washington

Technical Report No. 53

PLANKTON PIGMENT NOMOGRAPHS
by

A. C. Duxbury and C. S. Yentsch

Technical Report No. 54

THE LIGNIN FRACTION OF MARINE SEDIMENTS
by
Richard G. Bader

Office of Naval Research
Contract N8onr-520/III
Project NR 083-012

Reference 57-14
October 1956

Reference 57-15
December 1956

Clifford A. Barnes

For
RICHARD H. FLEMING
Executive Officer

The lignin fraction of marine sediments

RICHARD G. BADER

Abstract—The lignin in 31 surface samples of marine sediments from the Gulf of Maine and 26 samples from two cores collected in Puget Sound is an extremely variable fraction of their organic matter. In the two sediment cores, the loss of nonligninuous carbon is about six times that of lignin carbon. Apparently lignin is extremely stable in the marine environment.

Lignin is important from a geological standpoint. Knowledge of its concentration in sediments may permit palaeoecologic interpretations which are not possible by use of total organic carbon alone. It is potentially useful as a tool in studies of rates of sedimentation; but it must be determined whether it is dependent or independent of the deposition of inorganic particles.

The possible effects of lignin on carbon-nitrogen relationships and the base-exchange capacity of the organic matter in marine sediments are also discussed.

INTRODUCTION

LITTLE is known of the distribution of lignin in the marine environment. Its presence in marine sediments was first reported by TRASK (1932) and WAKSMAN (1933). BADER (1954) studied some surface concentrations of lignin in sediments from the Gulf of Maine.

Lignin is a very resistant organic complex, formed only by living plants (BERGENSTEIN, in BRAUNS, 1952) never by enzymatic or bacterial decay of other plant products. The decomposition of lignin is extremely slow (WAKSMAN and TENNY, 1926, 1927, 1929; BORUFF and BUSWELL, 1934; LEVINE *et al.*, 1935; BRAUNS, 1952, and others) and relatively constant under both aerobic and anaerobic conditions (ACHARYA, 1935, 1935a). In aerobic soils lignin is decomposed in traces or not at all (WAKSMAN and STEVENS, 1929, 1929a). It is the most resistant of all common plant materials. It is possible that in the microbiological decomposition of plant materials, lignin may act as an inhibitory factor (REGE, 1927; WAKSMAN and IYRE, 1932; FULLER and NORMAN, 1943) by the formation of stable lignoprotein complexes.

GOTTLIEB and HENDRICKS (1945) and others have attempted to isolate lignin from soils but were unable to secure definitely characterizable products. Methyl groups are apparently split off the lignin molecule (WAKSMAN and SMITH, 1934) and its cation exchange capacity increases (MILLER *et al.*, 1936). Despite the evidence for marked alteration, SCHOFFER and WELTE (1950) observed a striking similarity between the ultra-violet spectra of soil-lignin complexes and isolated lignin.

In the present investigation three general aspects were examined: (1) its distribution and variation in some marine sediments, (2) its stability in subsurface samples, and (3) its relationship to the remaining organic material.

METHOD OF ANALYSIS

Other organic compounds interfere with lignin determinations (WAKSMAN, 1933; NORMAN, 1937; BRAUNS, 1952) and must be removed. Lignin must then be determined indirectly because of its complexity and resistance to solution.

To remove the nonligninuous organic matter, an extraction and hydrolysis series was used. From 50 to 100 grams of dried sediments were crushed, powdered, and then treated with acetone in a

Soxhlet extraction apparatus for 48 hours. The sediment was then removed from the Soxhlet and refluxed in 95 per cent ethyl alcohol for 24 hours. The solvent was changed three times during this period in order to increase the efficiency of extraction. This was followed by a transfer of the sample to a fritted disk filter. The sediment was then vacuum filtered and thoroughly washed with fresh, hot, ethyl alcohol. The sample was then dried, removed from the filter and the filter was washed with 4 per cent HCl. The first hydrolysis was then accomplished by refluxing the sediment in 4 per cent HCl. After 24 hours, the sample was vacuum filtered and washed thoroughly with distilled water, transferred to 72 per cent H₂SO₄ and kept at 8°C for three hours. This solution was then diluted to 4 per cent H₂SO₄ and refluxed for four hours. Following vacuum filtration and washing with distilled water the samples were dried, powdered, and thus made ready for the determination of lignin carbon. The residual organic material after the H₂SO₄ treatment is termed lignin; this includes its transformation products as well as nitrogenous compounds such as synthesized microbial lignoprotein complexes. The carbon from this source shall thus be called lignin carbon; the remaining carbon is termed nonlignin carbon and represents that carbon removed by previous treatment.

Table 1. Surface sample data from are bounded by 44°05' to 44°28' N and 68° to 68°30' W

Sample No.	Depth (ft.)	Total carbon (%)	Lignin carbon (%)	Nonlignin carbon (%)	$\frac{\text{Lignin carbon}}{\text{Total carbon}} \times 100$
1	57	0.88	0.38	0.50	43.2
2	27	1.72	0.66	1.06	38.4
3	20	0.52	0.18	0.34	34.6
4	40	1.23	0.85	0.38	69.1
5	52	0.94	0.40	0.50	42.6
6	20	0.42	0.18	0.24	42.8
7	14	0.45	0.20	0.25	44.4
8	35	0.66	0.31	0.35	47.0
9	30	1.02	0.73	0.29	71.6
10	25	0.46	0.21	0.25	45.7
11	20	0.35	0.07	0.28	20.0
12	55	1.74	0.69	1.05	39.7
13	85	1.15	0.38	0.77	33.0
14	75	0.49	0.16	0.33	32.7
15	30	4.12	2.76	1.36	67.0
16	33	4.74	2.48	2.26	52.3
17	22	4.12	2.56	1.56	62.1
24	140	0.34	0.09	0.25	26.5
25	150	0.30	0.07	0.23	23.3
26	75	1.96	1.25	0.71	63.8
27	8	2.13	1.07	1.06	50.2
28	18	1.86	1.00	0.86	53.8
29	30	2.12	1.28	0.84	60.4
30	70	2.34	1.44	0.90	61.5
31	85	1.78	0.32	1.46	18.0
32	87	0.26	0.09	0.17	34.6
33	95	1.38	0.48	0.90	34.8
34	71	0.34	0.05	0.29	14.7
35	61	0.86	0.15	0.72	17.2
36	67	3.17	2.11	1.06	66.6
37	65	0.37	0.08	0.29	21.6
Mean			0.73	0.69	43.0
Standard deviation			0.25	0.15	16.8
Coefficient of variation			34.2	22.7	39.0

The total organic carbon and the lignin carbon were determined by a semimicro dry combustion method. The samples, dried to constant weight, were combusted in a measured volume of oxygen under controlled pressure. The evolved CO₂ was absorbed on Ascarite, determined gravimetrically and the per cent carbon computed. In determining the total organic carbon the sample was first

treated with warm, 4 per cent HCl to remove the carbonates. The use of such dilute HCl causes no loss of carbon from the organic matter. Carbon determinations by this method are reproducible to ± 0.03 per cent.

DISTRIBUTION AND RELATIVE STABILITY

The total organic carbon, lignin carbon, and the nonligninuous carbon was determined for the 31 surface samples of marine sediments off the coast of Maine in the vicinity of Mt. Desert Island. These samples were collected by means of a "snapper" sampler and a gravity coring tube (see Table 1). The wide range and relatively large coefficient of variation of lignin carbon in total carbon indicates that lignin does not have a constant ratio to the other organic constituents.

Core No. 169 from Puget Sound (Table 2) is primarily composed of inorganic particles of a relatively uniform size in the silt range (0.062-0.004 mm). Between the depths of 15 and 43 cm in the core, the particle size grades from silt to fine sand (0.125 mm). From 43 to 65 cm the particle size grades back into the silt range. The

Table 2. Subsurface sample data

Sample No. and location	Depth (cm)	Total carbon (%)	Lignin carbon (%)	Nonligninuous carbon (%)	$\frac{\text{Lignin carbon}}{\text{Total carbon}} \times 100$	
169 (250 ft.)	0	1.02	0.66	0.36	64.7	
	6	0.94	0.64	0.30	68.1	
	15	0.92	0.63	0.29	69.2	
	30	0.37	—	—	—	
	43	0.09	0.06	0.03	66.6	
	55	0.36	—	—	—	
	47°21.5'N 123°08.2'W	65	0.79	0.64	0.15	81.0
		85	0.76	0.62	0.14	81.6
		110	0.74	0.47	0.27	63.4
		115	0.74	0.69	0.05	93.2
125		0.79	—	—	—	
134		0.78	—	—	—	
140		0.77	0.60	0.17	77.9	
148	0.85	0.64	0.21	75.3		
221-C (200 ft.)	0	1.96	1.16	0.80	59.2	
	7	1.63	1.08	0.55	66.3	
	16	1.56	1.18	0.38	75.7	
	32	1.93	1.11	0.82	57.6	
	43	1.71	0.98	0.73	57.3	
	47°43.9'N 122°31.7'W	62	2.09	0.90	1.19	43.0
		74	2.38	1.50	0.88	63.0
		97	1.97	1.02	0.95	51.8
		115	2.27	1.08	1.19	47.6
		125	1.78	1.49	0.29	83.7
		150	2.00	1.37	0.63	68.5
		155	1.98	1.16	0.82	58.5

total organic carbon correspondingly decreases in this sandy layer reaching a minimum value at 43 cm. The remaining variations in organic content are not associated with particle-size changes. The maximum concentration of total carbon and non-ligninuous carbon occurs on the surface. With some minor variations, the per cent lignin carbon in the total carbon increases with depth.

Core No. 221-C from Puget Sound (Table 2) consists entirely of relatively uniform silt-sized particles. The fluctuations in organic content are not associated with

particle-size variations. The maximum concentration of total organic carbon is at 74 cm. For the immediate surface region, the per cent lignin carbon of the total carbon increases with depth, but below 16 cm this characteristic is not evident.

It appears that in the immediate surface sediments (0 to 6-7 cm) of both cores (Fig. 1) the lignin carbon decreases less rapidly with depth than the nonligninuous carbon. Core No. 169 has a lignin carbon loss of 3.0 per cent and a nonligninuous carbon loss of 16.6 per cent in the top 6 cm. Core 221-C has a 6.8 per cent lignin carbon loss and nonlignin carbon loss of 52.5 per cent in the top 7 cm. If it be assumed that environmental conditions have been relatively constant in both localities during this period of decomposition and that the organic decrease is due to decomposition, the percentage of nonligninuous carbon loss is 5.5 to 7.7 times that of the lignin carbon. This indicates that the ligninuous fraction is more resistant to decomposition than the greater part of the other organic compounds. These data (Table 2, Fig. 1) indicate that changes in the lignin carbon in excess of 1 per cent or 2 per cent per cm are due to supply changes, i.e., the lignin carbon loss due to decomposition is less than 1 per cent to 2 per cent of the total amount of lignin carbon.

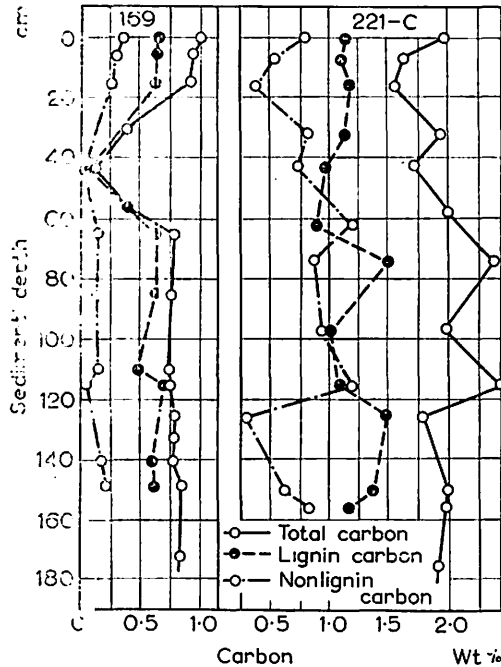


Fig. 1. Vertical distribution of total organic carbon, lignin carbon, and nonligninuous carbon in two cores of Puget Sound sediments. The change in total, lignin, and nonlignin carbon in core 169 between 15 and 65 cm is a reflection of an inorganic particle size change, i.e. a sand layer. The remaining variations in core 169 and all of those in core 221-C are not related to particle size changes.

SOURCE OF ORGANIC MATTER

Although the greater part of the organic matter in marine sediments may be derived from the decomposition of planktonic organisms (SVERDRUP *et al.*, 1942) in regions of significant river drainage a considerable fraction may be derived from the land (GRIPENBERG, 1934). Since all analyses of ligninuous material have been made

on samples obtained nearshore (Tables 1 and 2; TRASK, 1932; WAKSMAN, 1933), much, if not all, of this fraction too may have been derived from the land. Indeed, it has not yet been demonstrated that marine algae have lignin-like complexes similar to those of terrestrial algae (WAKSMAN, 1936). Hence, no definite conclusions can yet be made as to the actual source of the ligninous matter under consideration here.

SEDIMENTATION

The amount of organic matter included in marine sediments is related to the rate of inorganic sedimentation as well as to organic supply and decomposition. Variations in concentrations of organic matter can be explained by the following means:

(1) A high rate of sedimentation of inorganic material would result in diluting the simultaneously deposited organic matter and *vice versa*.

(2) A low rate of sedimentation would increase the time that the organic fraction remains in the upper oxidizing zone of the sediments. It would thus have a longer period of active decomposition.

(3) A decrease in the supply of organic material would cause a corresponding decrease in the sediments, and *vice versa*.

(4) Environmental changes not effecting the supply or rate of sedimentation of organic material may alter the rate of microbiological decomposition.

The data from Core 221-C (Fig. 1) shows a decrease in total organic carbon from 74 to 43 cm. This could be explained on the basis of any of the four mechanisms listed above. One might conclude that this segment existed in an environmental situation that changed continually and gradually with time. However, the lignin carbon concentration decreases appreciably from 74 to 62 cm, as the nonligninous carbon increases. From 62 to 43 cm the lignin carbon increases slightly although the nonlignin carbon decreases markedly. Similar situations occur between 97 and 74 cm and 150 to 155 cm. These observations of the ligninous portion indicate that there was not a constantly and gradually changing environment over these depth ranges; other factors must be involved.

CARBON-NITROGEN RATIO

The wide variation in the lignin content of marine sediments and its resistance to decomposition may explain the wide variations in carbon-nitrogen ratios in marine sediments (GRIPENBERG, 1934; MOHAMED, 1949; ARRHENIUS, 1950; BADER, 1955). Since lignin may inhibit organic decomposition and may be associated with the microbiological formation of stable lignoprotein complexes (REGE, 1927; WAKSMAN and IYER, 1932; FULLER and NORMAN, 1943), sediments with a high lignin content have the potentiality of "trapping" more nitrogen per unit of organic material than sediments with low lignin concentrations, since nitrogen associated with lignin would be less mobile than that in the remaining organic compounds.

BASE-EXCHANGE CAPACITY

Discussions of base exchange in marine sediments have been confined to the clay-water system (KELLY, 1955) and little is known of the effect of organic material in this connection.

The organic fraction of soils has approximately seven to eight times the exchange capacity of the mineral fraction (TURNER, 1932; GEDROIZ, 1933). The base exchange

capacity of soil organic matter accounts for as much as 61 per cent of the total capacity (MITCHELL, 1932; WILLIAMS, 1932). In a podsol soil 87 per cent of the exchange capacity was due to the organic constituents (BARSHAD and ROJAS-CRUZ, 1950). In a soil containing 4 per cent organic matter, well within the range found in continental-shelf sediments, 40 per cent of the total exchange capacity has been attributed to this organic fraction. Lignin complexes are capable of irreversibly adsorbing NH_3 and other bases (WEDEKIND and GARRE, 1928).

Although free lignin has a very low exchange capacity, when it is combined with protein to form a complex, the increase in base exchange capacity far exceeds the sum of the two and closely approximates the exchange capacity of the total humus (WAKSMAN and IYER, 1932). These data indicate that the lignoprotein fraction may be responsible for much of the base-exchange capacity in the organic matter of marine sediments. The contribution to the total exchange capacity by the organic matter in the sediments should vary markedly according to the amount of lignin complexes present. Hence, organic matter in general and lignin in particular may be important factors in the early diagenetic processes in marine sediments.

SUMMARY

(1) The total lignin content and its ratio to the total organic material in marine sediments is extremely variable.

(2) Lignin is an organic complex displaying extreme resistance to microbiological decomposition. The loss of nonlignin carbon is approximately six times greater than the lignin carbon loss. Lignin carbon changes which exceed 1 per cent to 2 per cent per cm of the total may thus be ascribed to changes in the amount originally supplied to the sediments.

(3) Concentration of carbon from lignin sources in marine sediments should give further information for palaeoecology.

(4) The concentration of lignin in marine sediments may be associated with variations in carbon-nitrogen ratios, due to its variability, stability, and the formation of nitrogen-bearing lignin complexes.

(5) When investigating the physicochemical system in marine sediments the effect of lignin and lignin complexes on this system should be considered, since studies of soils indicate that this fraction accounts for most of the base-exchange capacity attributed to organic materials.

Acknowledgements—This study was supported by the Office of Naval Research Contract N8onr-520/111. The writer also wishes to thank CLIFFORD A. BARNES and HERBERT J. FROLANDER of the University of Washington as well as DONALD W. HOOD and JOHN P. BARLOW of Texas A. & M. for their helpful suggestions.

Contribution No. 200 from Department of Oceanography, University of Washington, Seattle, Washington. Author's present address: Department of Oceanography, A. & M. College of Texas, College Station, Texas.

REFERENCES

- ACHARYA C. N. (1935) Studies on the anaerobic decomposition of plant materials, I: Anaerobic decomposition of *Oryzo sativa*. *Biochem. J.* **29**, 528-541.

- ACHARYA C. N. (1935a) Studies on the anaerobic decomposition of plant materials, III: Comparison of the course of decomposition of rice straw under anaerobic, aerobic and partially aerobic conditions. *Biochem. J.* **29**, 1116-1120.
- ARRHENIUS G. (1950) Carbon and nitrogen in subaquatic sediments. *Geochim. et Cosmochim. Acta* **1**, 15-21.
- BADER R. G. (1954) The role of organic matter in determining the distribution of Pelecypods in marine sediments. *J. Mar. Res.* **13**, 32-47.
- BADER R. G. (1955) Carbon and nitrogen relations in surface and subsurface marine sediments. *Geochim. et Cosmochim. Acta.* **7**, 205-211.
- BARSHAD I. and ROJAS-CRUZ L. A. (1950) A pedologic study of podsol soil profile from the equatorial region of Colombia, South America. *Soil Sci.* **70**, 221-236.
- BORUFF C. S. and BEESWELL A. M. (1934) The anaerobic fermentation of lignin. *J. Amer. Chem. Soc.* **56**, 885-888.
- BRAUNS F. E. (1952) *The Chemistry of Lignin*. Academic Press Inc., New York, 808 pp.
- FULLER W. H. and NORMAN A. G. (1943) Cellulose decomposition by aerobic mesophilic bacteria from soil, III: The effect of lignin. *J. Bact.* **46**, 291-297.
- GEDROIZ K. K. (1933) Influence of hydrogen peroxide upon soil. *Proc. 2nd Intern. Cong. Soil Sci.* (1930) **2**, 41-70.
- GOTTLIEB S. and HENDRICKS S. B. (1945) Soil organic matter as related to newer concepts of lignin chemistry. *Soil Sci. Soc. Amer. Proc.* **10**, 117-125.
- GRIPENBERG STINA (1934) A study of the sediments of the North Baltic and adjoining seas. *Fennia* **60**, 231.
- KELLY W. P. (1955) Base exchange in relation to sediments, In Recent Marine Sediments by TRASK, P. D. *Sp. Pub. Soc. Econ. Paleon. and Min.* **4**, 454-465.
- LEVINE M., NELSON G. H., ANDERSON D. Q. and JACOBS P. B. (1935) Utilization of Agricultural wastes, I: Lignin and microbial decomposition. *J. Ind. Engin. Chem.* **27**, 195-200.
- MILLER H. C., SMITH F. B. and BROWN P. E. (1936) The base exchange capacity of decomposing organic matter. *J. Amer. Soc. Agron.* **24**, 256-274.
- MITCHELL J. (1932) The origin, nature and importance of soil organic constituents having base exchange properties. *J. Amer. Soc. Agron.* **24**, 256-274.
- MOHAMED A. F. (1949) The distribution of organic matter in sediments from the Northern Red Sea. *Amer. J. Sci.* **247**, 116-127.
- NORMAN A. G. (1937) *The Biochemistry of Cellulose, the Polyuronides, Lignin, etc.* Clarendon Press, Oxford, 232 pp.
- REGE R. D. (1927) Biochemical decomposition of cellulosic materials, with special reference to the action of fungi. *Ann. Appl. Biol.* **14**, 1-44.
- SCHAEFFER F. and WELTE E. (1950) Die Anwendung der Absorption spektrographic in der Humus Forschung. *Ztschr. Pflanzenernahr. Dung Bodenk.* **48**, 250-263.
- SVERDRUP H. U., JOHNSON M. W. and FLEMING R. H. (1942) *The Oceans*. Prentice-Hall Inc., New York, 1087.
- TENNY F. G. and WAKSMAN S. A. (1929) Composition of natural organic materials and their decompositions in the soil: IV. *Soil Sci.* **28**, 55.
- TRASK P. D. (1932) *Origin and Environment of Source Sediments of Petroleum*. Gulf Pub. Co., Houston, 323 pp.
- TURNER P. E. (1932) An analysis of factors contributing to the determination of saturation capacity in some tropical soils. *J. Agri. Sci.* **22**, 72-91.
- WAKSMAN S. A. (1933) On the distribution of organic matter in the sea bottom and the chemical nature and origin of marine humus. *Soil Sci.* **36**, 125-147.
- WAKSMAN S. A. and STEVENS K. R. (1929) Processes involved in the decomposition of fossilized wood. *J. Amer. Chem. Soc.* **51**, 1187-1196.
- WAKSMAN S. A. and STEVENS K. R. (1929a) Contribution to the chemical composition of peat: V. *Soil Sci.* **28**, 315-340.
- WAKSMAN S. A. and TENNY F. G. (1926) On the origin and nature of soil, organic matter of soil "humus": IV. *Soil Sci.* **22**, 395-406.
- WAKSMAN S. A. and TENNY F. G. (1927) The composition of natural organic materials and their decomposition in the soil, II: Influence of age of plant upon rapidity and nature of its decomposition. *Soil Sci.* **24**, 317-333.
- WAKSMAN S. A. and IYRE K. R. N. (1932) Contribution to our knowledge of the chemical nature and origin of humus, I: On the synthesis of the "humus nucleus." *Soil Sci.* **34**, 43-71.

- WAKSMAN S. A. and SMITH H. W. (1934) Transformation of the methoxyl group in lignin, in the processes of decomposition of organic residues by marine organisms. *J. Amer. Chem. Soc.* **56**, 1225-1229.
- WAKSMAN S. A. (1935) Chemical nature of soil organic matter, methods of analysis, and the role of micro organisms in its formation and decomposition. *Soil Sci.* **40**, 347-365.
- WEDEKIND VON E. and GARRE G. (1928) Die Kolloidnatur der Ligninsäure und ses sog Kasseler Brauns; ein Vergleich. *Kolloid Ztschr.* **44**, 205-212.
- WILLIAMS R. (1932) The contributions of clay and organic matter to the base exchange capacity of soils. *J. Agri. Sci.* **22**, 845-851.

Department of Oceanography
University of Washington

UNCLASSIFIED TECHNICAL REPORT DISTRIBUTION LIST

- | | |
|--|--|
| 3 Chief of Naval Research
Department of Navy
Washington 25, D. C.
Attn: Code 446 (1)
463 (1)
466 (1) | 3 Chief, Bureau of Ships
Department of the Navy
Washington 25, D. C.
Attn: Code 312 (1)
320 (1)
845 (1) |
| 1 Commanding Officer
Office of Naval Research Branch Office
346 Broadway
New York 13, New York | 1 Chief, Bureau of Yards and Docks
Department of the Navy
Washington 25, D. C. |
| 1 Commanding Officer
Office of Naval Research Branch Office
John Crerar Library Building
86 East Randolph Street
Chicago 1, Illinois | 1 Chief of Naval Operations
Department of the Navy
Washington 25, D. C.
Attn: Op-533D |
| 1 Commanding Officer
Office of Naval Research Branch Office
1030 East Green Street
Pasadena 1, California | 1 Commander
Naval Ordnance Laboratory, White Oak
Silver Spring 19, Maryland |
| 1 Commanding Officer
Office of Naval Research Branch Office
1000 Geary Street
San Francisco 9, California | 1 Commanding Officer
U. S. Navy Mine Countermeasure Station
Panama City, Florida |
| 3 Commanding Officer
Office of Naval Research Branch Office
Navy #100, Fleet Post Office
New York, New York | 1 Commanding Officer
U. S. Navy Underwater Sound Laboratory
New London, Connecticut |
| 2 Office of Naval Research
Geophysics Branch
Washington 25, D. C.
Attn: 416 | 2 Department of Aerology
U. S. Naval Post Graduate School
Monterey, California |
| 1 Office of Naval Research
Resident Representative
University of Washington
Seattle 5, Washington | 3 Director
U. S. Navy Electronics Laboratory
San Diego 52, California
Attn: 2230 |
| 2 Chief, Bureau of Aeronautics
Department of the Navy
Washington 25, D. C.
Attn: PH 41 (1)
AY -3 (1) | 6 Director
Naval Research Laboratory
Washington 25, D. C.
Attn: Technical Services Information
Officer |
| | 8 Hydrographer
U. S. Navy Hydrographic Office
Washington 25, D. C.
Attn: Division of Oceanography |

- 1 Project Arowa
U. S. Naval Air Station
Building R-48
Norfolk, Virginia
- 1 Superintendent
U. S. Naval Academy
Annapolis, Maryland
- 5 Armed Services Technical Information
Center
Documents Service Center
Knott Building
Dayton 2, Ohio
- 1 Assistant Secretary of Defense
for Research and Development
Pentagon Building
Washington 25, D. C.
Attn: Committee on General Sciences
- 1 Chief, Air Weather Service
Department of the Air Force
Washington 25, D. C.
- 1 Chief
Armed Forces Special Weapons Project
P. O. Box 2610
Washington, D. C.
- 2 Chief, U. S. Weather Bureau
2400 M Street N. W.
Washington 25, D. C.
Attn: Dr. H. Wexler
- 1 Commandant (OAO)
U. S. Coast Guard
Department of the Treasury
Washington 25, D. C.
- 1 Commanding General
Research and Development Division
Department of the Air Force
Washington 25, D. C.
- 1 Commanding General
Research and Development Division
Department of the Army
Washington 25, D. C.
- 1 Commanding Officer
Cambridge Field Station
230 Albany Street
Cambridge 30, Massachusetts
Attn: CRHSL
- 1 Director
U. S. Coast & Geodetic Survey
Department of Commerce
Washington 25, D. C.
- 2 Director, U. S. Fish & Wildlife Service
Department of the Interior
Washington 25, D. C.
Attn: Dr. L. A. Walford
- 1 National Research Council
2101 Constitution Avenue
Washington 25, D. C.
Attn: Committee on Undersea Warfare
- 1 Office of Technical Services
Department of Commerce
Washington 25, D. C.
- 1 U. S. Army Beach Erosion Board
5201 Little Falls Road N. W.
Washington 16, D. C.
- 2 U. S. Fish & Wildlife Service
Pacific Oceanic Fishery Investigation
P. O. Box 3830
Honolulu, T. H.
Attn: Librarian (1)
T. S. Austin (1)
- 1 U. S. Fish & Wildlife Service
Scripps Institution of Oceanography
La Jolla, California
- 1 U. S. Fish & Wildlife Service
Woods Hole, Massachusetts
- 1 U. S. Waterways Experiment Station
Vicksburg, Mississippi
- 1 Allen Hancock Foundation
University of Southern California
Los Angeles 7, California

- | | |
|--|--|
| 1 Bingham Oceanographic Laboratories
Yale University
New Haven, Connecticut | 1 Director
National Institute of Oceanography
Wormley, Near Godalming
Surrey, England |
| 1 Department of Conservation
Cornell University
Ithaca, New York
Attn: Dr. J. C. Ayers | 2 Director
Scripps Institution of Oceanography
La Jolla, California |
| 1 Department of Engineering
University of California
Berkeley, California | 2 Director
Woods Hole Oceanographic Institution
Woods Hole, Massachusetts |
| 1 Department of Meteorology & Oceanography
College of Engineering
New York University
University Heights, New York 53, N. Y.
Attn: Dr. W. J. Pierson | 1 Dr. Wayne V. Burt
Oregon State College
Corvallis, Oregon |
| 1 Department of Zoology
Rutgers University
New Brunswick, New Jersey
Attn: Dr. H. K. Haskins | 1 Head, Department of Oceanography
Texas A & M College
College Station, Texas |
| 1 Director
Bermuda Biological Station for Research
St. George's, Bermuda, B. W. I. | 1 Hudson Laboratories
Columbia University
145 Palisades Street
Dobbs Ferry, New York |
| 1 Director, Chesapeake Bay Institute
Box 42, R. F. D. #2
Annapolis, Maryland | 1 Institute of Oceanography
University of British Columbia
Vancouver, British Columbia, Canada |
| 1 Director, Hawaii Marine Laboratory
University of Hawaii
Honolulu, T. H. | 2 Project Officer
Laboratory of Oceanography
Woods Hole, Massachusetts |
| 1 Director
Lamont Geological Observatory Torrey Cliff
Palisades, New York | |
| 1 Director, Marine Laboratory
University of Miami
Coral Gables, Florida | |
| 1 Director
Narragansett Marine Laboratory
Kingston, Rhode Island | |