

1 Crab on a stick: assessing spatial variation in predator community and predation rate using
2 tethered megalopae and underwater video

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44 **Abstract**

45 Planktonic larval development, common in many benthic marine invertebrates, is associated with
46 both costs and benefits relative to benthic larval development. Costs of being in the plankton
47 include increased susceptibility to advection away from habitat and starvation. Potential benefits
48 include reduction in predation for pelagic larvae. Few studies have measured predation rates on
49 larvae in the field. Even fewer have directly observed larval predation events, documented the
50 specific predator, mode of predation, and intervals between predator attacks. In this study we
51 investigated benthic and pelagic differences in predator assemblage and predation interval on
52 megalopae of the Dungeness crab, *Metacarcinus magister*. To observe the predation events, we
53 combined methods of tethering larvae with automated motion analysis of underwater camera
54 images from daytime deployments off the dock at Friday Harbor Laboratories, Friday Harbor,
55 WA. We measured number of predation events, the interval in which those events happened, the
56 species of predator, and of other non-predator species that comprised each visible community.
57 Predation was higher on the benthos than the surface, with more total strikes and a shorter
58 interval to predation. Only one predator species was seen during the trials, *Artemia harringtoni*.
59 These results are consistent with previous studies, and support the novel combination of methods
60 to better inform predation interactions of communities and their relative importance on each
61 species.

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69 **Introduction**

70 Larval development in the plankton is common amongst benthic marine invertebrates
71 (Marshall et al. 2012). Having a planktonic life stage incurs new risks compared to an entire life
72 cycle in the benthos; these risks include advection away from habitat, starvation before
73 metamorphosis, and predation on larvae as they transition to a pelagic habitat. Conversely, a
74 planktonic stage can also provide additional benefits in the forms of escape from predation,
75 additional food, and dispersal (Strathmann et al. 2002). Although a planktonic stage has been
76 hypothesized to reduce predation on larvae, absolute mortality rates can still be substantial
77 during the larval stage, which have major implications for the rates of juvenile recruitment back
78 to the benthic environment (Strathmann 1985, Morgan 1995).

79 Reviews of both field and laboratory studies demonstrated predation on larvae to be a
80 potentially significant source of mortality (Morgan, 1995; Allen and Vaughn, 2010). Estimates
81 of larval mortality due to predation in the field have ranged from 0-100% (Rumrill, 1990;
82 Johnson and Shanks, 2003; Allen and McAllister, 2007). Possible explanations include
83 differences between species; differences in methods of measuring predation rates; and
84 differences in geography, seasonality, and other spatiotemporal factors. Another possible
85 explanation for large variance in predation rates could be that predation varies spatially over very
86 short length scale, *i.e./e.g.* across water column depth and shallow subtidal environments.
87 Predation can be substantially higher in near shore environments compared to offshore habitats
88 (Morgan, 1990), and in another study, predation rates in tethered megalopae were shown to
89 differ significantly between the benthos and the water column (Allen and McAllister 2007).

90 A lack of direct methods for observing larval predation has been an obstacle for previous
91 studies. Methods, such as tracking a known amount of larvae, monitoring a volume of water over
92 time (Johnson and Shanks, 2003), and quantifying the loss of tethered larval prey (Allen and
93 McAllister, 2007; D’Alessandro and Sponaugle, 2011; Kerr et al. 2014) can provide invaluable
94 estimates of predation rates. However, these studies do not establish exactly when a predation
95 event occurs and therefore, are unable to resolve a precise attack interval on prey items. Indirect
96 methods quantify predation loss after a set amount of time, overestimating the interval to a
97 predation event, resulting in an underestimate of predation rate. If the attack interval is known,
98 predation rates can be more accurate.

99 The other issue these methods have is discerning the predator assemblage for the study
100 area. Predator assemblages can be inferred from environmental surveys (Kerr et al. 2014) or
101 from attaching hooks to tethered larvae (Fernandez et al. 1993). But surveys might miss
102 uncommon predators, and not all predators can be captured by hooks. Alternatively, stable
103 isotope or gut analysis of potential predators could assess the predator assemblage, but attack
104 interval cannot be found and choice of focal predators can bias the results.

105 In this study, we investigated spatial variation in the predator assemblages and the
106 predation interval on megalopae of the Dungeness crab, *Metacarcinus magister* (Dana, 1852). To
107 determine the species assemblage and predation interval at both the benthic and planktonic level,
108 we used tethering methods in conjunction with a novel underwater camera housing that enabled
109 direct observation of predation on the megalopae. Based on Allen and McAllister (2007), we
110 predicted that attack interval would be quicker on the benthos compared to the plankton. We also
111 predicted that the species assemblage would differ between the benthos and the plankton.

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113 **Methods**

114 To examine the predator community of *M. magister* megalopae, we tethered megalopae
115 in close proximity to underwater camera housings and documented predation events over 2 hour
116 intervals, during daylight from 30 July 2014 to 3 August 2014. We also recorded predator
117 assemblage differences between the benthos and the surface.

118 **Study site**

119 We conducted our study off the docks at Friday Harbor Laboratories, Friday Harbor, WA
120 (N 48° 32' 46" N, W 123° 0' 46"). This dock is protected by a floating breakwater, but boat
121 traffic causes wakes to the floating dock. Pier pilings are spaced on either side roughly 3.3m
122 apart from shore to sea and roughly 5m apart perpendicular to shore. The benthic deployment
123 site is a soft sediment bottom with patchy kelp as well as debris algae and eelgrass transported by
124 tidal exchanges. The depth to the benthos is 2.13m when tide is at 0m MLLW.

125 **Camera housing**

126 We used a 4.50m x 5.08cm and a 1.80m x 5.08cm PVC tube for the benthic and surface
127 camera housings respectively. With PVC cement and silicone sealant, we glued one end of each
128 PVC tube to half of a coupler and an L-shaped elbow to the other half of the coupler and screwed
129 the couplers together to create a watertight seal. 78.0mm x 80.0mm x 5.0mm pieces of acrylic
130 plastic were glued to the open ends of the L-shaped elbows using epoxy. A 5.83kg weight was
131 tied to the 4.5m housing, and an 8.5kg weight was used for the 1.8m housing, to provide negative
132 buoyancy and stabilization of the camera and the prey.

133 **Camera**

134 We used modified web cameras (Logitech QuickCam Pro 9000) for the
135 experiment, which slid into the PVC housing until they reached the elbow, facing out of the

136 acrylic window. The camera was modified by altering the focal distance to be approximately
137 5cm. The cameras were powered by a laptop connected with USB extension cables to reach out
138 of the PVC tubes. iSentry and iCamSource, the applications we used, were designed to capture
139 pictures from the external webcams when motion was detected in the field of view. Both
140 applications were set to the highest sensitivity setting and recorded images at 5 frames per
141 second (fps) when triggered. The pictures from each deployment were then assembled into a
142 movie using Time Lapse Assembler. We used the movies to note when the first predation events
143 occurred and identified all the animals that approached the megalopae. For the purposes of this
144 study, we defined a predation event is as an attempt by a predator to consume the megalopa,
145 whether or not the attack was successful.

146 In order to steady the cameras within the housings, we attached them to separate 2x4
147 blocks of wood using U-Bolts. The 2x4x12 wood block for the benthic camera housing was
148 nailed to pilings that were attached to the substrate, while the 2x4x6 wood block for the surface
149 camera housing was nailed to the floating dock. This allowed the benthic camera to remain near
150 the bottom and the surface camera to always be 1.05m below the water line even as the tides
151 fluctuated.

152 **Tethering megalopae**

153 Megalopae were collected off the Friday Harbor Laboratories docks at night using a
154 nightlight and glass jars. The tethers consisted of a 4cm length of 0.85mm diameter fishing line
155 with a 1cm Nitex thread superglued, to it, adjusted to place the megalopa within the focal plane
156 and the field of view. The fishing line provided rigidity to the tether, allowing the megalopae to
157 stay within the field of view of the cameras, while the Nitex thread was flexible enough to allow
158 the megalopae some movement. Before tethering, we measured the carapace length of the

159 megalopae here defined as from the eye to the start of the abdomen to the nearest 0.5mm. We
160 superglued the megalopae onto the Nitex thread and attached the tether to the acrylic using
161 binder clips.

162 **Capturing footage**

163 Housings with tethered megalopae were placed into the water for 2 hours, consistent with
164 the sampling intervals of Allen & McAllister (2007). Megalopae were replaced, even if not
165 consumed before the beginning of the next trial. Trials started at approximately 8am and ended at
166 approximately 9pm each day, when light levels became too low for photography.

167 **Control**

168 As a positive control, we crushed 17 recently molted *M. magister* juveniles, released
169 them under the docks, and tracked their trajectories until they were either advected out of view or
170 consumed by a predator. If predation by surface predators is typically high, but these predators
171 were deterred by the presence of the camera housings, freely floating crushed *M. magister* should
172 still be consumed. All of the *M. magister* sank out of sight without any predation event by
173 potential predators.

174 **Results**

175 Out of the 19 trials on the benthos, 8 predation events occurred all with *Artemius*
176 *harringtoni* as the predator. In contrast, none of the tethered megalopae were attacked near the
177 surface (Figure 1). Using a binomial logistic regression, there is a significant difference between
178 the proportion of predation events between surface and benthos ($X^2 = 12.781$, $p = 0.0003$).

179 Because we did not observe predation events on the surface, we are unable to compare
180 the predation intervals, defined here as the time from the introduction of the megalopa until it
181 was attacked, across treatment levels. However, we calculated upper and lower bounds for attack

182 rates in both the benthic and surface treatments (Table 1) If a megalopa was attacked on the
183 benthos, the predation interval averaged 50.11 minutes +/- 21.79 minutes (Table 1).

184 Our cameras also allowed us to identify other in the field. With the exception of one
185 ctenophore (*Pleurobrachia* sp.) found on the surface, all of the other potential predators were
186 observed near the benthos (Table 2). *A. harringtoni* and *Hexagrammos* spp. were the most
187 common fish recorded, but several crustaceans, such as cancrid crabs and *Pugettia* sp., were also
188 observed. Since we observed multiple individuals of the same species in one trial, it is possible
189 that the animals we documented were not independent of each other. The lack of independence
190 would violate statistical assumptions needed to determine if certain organisms were more
191 common than others.

192 **Discussion**

193 Our results support the findings of Allen and McAllister (2007) that predation of *M.*
194 *magister* megalopae is significantly higher in the benthos compared to the surface. Our estimate
195 of the percent of megalopae lost per minute, 0.35%, at the benthos was consistent with the
196 findings of Allen and McAllister (2007), 0.29%. Both study suggest that, for megalopae and
197 possible other similarly large and well-protected larvae, benthic predation in multiple sites is
198 significantly higher than at the surface. The absence of predation at the surface also corresponds
199 to the low predation in the plankton observed by Johnson and Shanks (2003). A limitation of our
200 method came from the surface camera being fixed onto a floating dock allowing for disturbances
201 from people being on the dock to potentially scare predators away. The results of our positive
202 control hint that predation is still low on the surface even without a camera housing. Tethering
203 megalopae may lead to an overestimate of predation rates because it precludes effective
204 responses from predators. Similarly, a tethered megalopa could have a lower encounter rate with

205 predators since it has relatively low motion with respect to the water and benthos compared to its
206 normal swimming speed of 4.2cm/s (Jacoby 1982). Since 100% of the predation events involved
207 *Artemia harringtoni* (Table 1), we concluded that it is the primary predator of megalopae at this
208 site during the day.

209 **Potential predator assemblage**

210 In addition to identifying the predator species on the megalopae, the camera allowed us to
211 observe other organisms in the different habitats. Data from the surface camera showed that
212 potential predators were not abundant near the top of the water column. This supports our control
213 where crushed *M. magister* deployed near the surface did not elicit any predation events despite
214 the absence of a camera housing.

215 The benthic camera, however, recorded several other species of fish and crustaceans that
216 could have been potential predators of megalopae. Various species of crabs and fish came into
217 close contact with the megalopae, perhaps attracted by the L-shaped elbow which resembled a
218 potential shelter, but made no attempts to consume the megalopae. While it would be
219 unreasonable to conclude that these species do not consume megalopae, their relatively frequent
220 occurrence on the benthos and the fact that only *Artemia harringtoni* preyed on megalopae
221 imply that these other species might not be major predators of megalopae. Our observation
222 contrasts with other studies, in which cannibalism from juvenile crabs was a significant source of
223 mortality (Fernandez et al. 1993; Moksnes et al. 1997). Interestingly, Fernandez et al. (1993) also
224 found that another species of sculpin, *Leptocottus armatus*, was the only predator in their
225 tethering experiments using *M. magister* megalopae at Gray Harbor estuary near the southern
226 Washington coast. This suggests that this group of fishes is an important, perhaps the most
227 important, factor determining the relative cost of the benthic environment as habitat for larval

228 development. In sites that lack sculpins a follow up study is needed to investigate whether other
229 types of predators assume the role of larval predators or whether those habitats are favorable for
230 larval development.

231 The conventional explanation for vertical migration rests upon the supposition that
232 predation risk for zooplankton is highest during the day (Lampert 1989). While we do not have
233 data on predation rates and intervals for night, we are still able to conclude that, under the
234 limitations of our experimental approach, the benthos appears to be more dangerous for
235 megalopae during the day than the surface. It is possible that other explanations for vertical
236 migration, such as aversion to UV light and reducing metabolism during the day, are driving the
237 movement of zooplankton.

238 Kerr et al. (2014) proposed another explanation for their observed lack of reduction in
239 predation between day and night. Although large zooplankton may face substantial predation risk
240 in the water column during the day, predation risk could actually increase at night since
241 nocturnal planktivorous fish target larger prey items (Holzman and Genin 2003). Kerr et al.
242 (2014) noticed a shift in predation risk when *Artemia franciscana* grew into adults of 3.7mm in
243 length. The megalopae we used had carapace width between 4 and 5 mm, so it is plausible that
244 they face similar predation risks as adult *A. franciscana*. This explanation of similar nighttime
245 predation to daytime predation for large zooplankton does not explain the lack of predation
246 events near the surface for our megalopae.

247 The results from this study support the idea proposed by Strathmann et al. (2002) that the
248 planktonic stage is an adaptation for larvae to escape predation from the benthos since we
249 observed a significant reduction in predation in the water column. Identifying the components of
250 mortality and how they change spatially can be important for this commercially valuable species.

251 Further observations of predation on megalopae at night, with infrared lighting, are needed to
252 fully understand how predation rates differ between surface and the benthos. Additional studies
253 at sites outside of the Friday Harbor Lab dock would aid in expanding the scope of the study and
254 better the understanding of surface predation rates.

255 With the use of this novel combination of methods, we were able to support previous
256 findings who found low predation rates on the surface (Johnson and Shanks, 2003) and increased
257 predation rates on the benthos (Allen and McAllister 2007), determine predator assemblage, as
258 well as attain more precise estimates of predation intervals on megalopae. By directly observing
259 predation events with underwater camera housings, we were able to obtain accurate estimates of
260 predation intervals and document a predator assemblage. Better estimates of predation intervals
261 allowed us to calculate improved predation rates, which have implications for population
262 dynamics. Recognizing the major predators on larvae refines our understanding of larval food
263 web ecology.

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322 Table 1. Table presenting the number of predation events, the predator species for each
 323 event, the predation interval, the % loss per minute for the two different depths, and the
 324 95% confidence interval for attack rate.
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Depth	Number of predation events	Predator species	Predation Interval (minutes)	% Loss per Minute	95% Confidence Interval for Attack Rate (# of attacks per second)
Surface	0	N/A	N/A	0	0 to 1.7e-5
Predation	8	<i>Artedius harringtoni</i>	50.11 +/- 21.79	0.034231921	3.3e-5 to 1.4e-4

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327 Table 2. A list of organisms found within the field of view of the camera during a trial that
328 did not consume a megalopa. An observation is defined when an organisms enters the field
329 of view until it leaves.

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Species observed near the benthos	Number of observations
<i>Hexagrammos stelleri</i>	4
<i>Hexagrammos</i> sp.	6
<i>Artedius harringtoni</i>	11
<i>Chirolophis</i> sp.	4
<i>Pholis laeta</i>	4
<i>Embiotoca lateralis</i>	3
<i>Pugettia</i> sp.	1
Embiotocidae	4
Cancriidae	4
Shrimp	4
<i>Pleurobrachia</i> sp. (surface)	1

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333 Figure 1. The proportion of predation events between megalopae tethered to the benthos
334 and those tethered near the surface ($X^2 = 12.781$, $p = 0.0003523$)
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