

Investigating the molecular basis of notochord loss in *Molgula occulta*
via transcriptome sequencing

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BEACON Research Fellowship 2011
Summer 2011

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Introduction

Ascidian tunicates, commonly known as ‘sea squirts’, are sessile hermaphroditic filter-feeders found in marine environments (Huber *et al.*, 2000). Tunicates are *urochordates* (i.e. ‘tail’ chordates) because they possess the defining chordate structure, the notochord, in a larval tail that is reabsorbed upon metamorphosis. This tail is believed to benefit progeny by facilitating motility, which in turn permits a wider variety of anchoring environments and a higher probability of finding favorable settling conditions (Huber *et al.*, 2000). Despite the utility of this adaptation the larval notochord has been independently lost several times among the ascidians (Huber *et al.*, 2000). These anural, or tailless, tunicates occur with the highest frequency in the Molgulidae family (Huber *et al.*, 2000).

This study uses a newly sequenced transcriptome of closely related molgulid ascidians to evaluate notochord-specific gene expression in the anural tunicate *Molgula occulta*, the tailed tunicate *Molgula oculata* and *M. occulta* X *M. oculata* hybrids. Highly conserved genes expressed in the notochords of the urodele species *Ciona intestinalis* and *Molgula tectiformis* were examined for expression in *M. occulta*, *M. oculata* and *M. occulta* x *M. oculata* hybrids via transcriptome analysis. The results of this study may further elucidate the expression patterns essential to notochord development and implicate the nature of the genes involved in the evolution of taillessness. This study may also facilitate creating a clearer tunicate phylogeny through phylogenetic analyses that assesses the conservation of these key developmental genes in a Molgulid genome.

One compelling gene identified in the *C. intestinalis* and *M. tectiformis* transcriptomes is *prickle*, a gene family that plays a pivotal role in the planar cell-polarity

(PCP) pathway of Wnt-signaling. Studies in mice and *Drosophila* have shown that the PCP pathway is involved in coordinating cell movements during development in both vertebrates and invertebrates (Tao *et al.*, 2009). Within *C. intestinalis* and *C. savignyi*, *prickle* is necessary for notochord intercalation and is most highly expressed in notochord cells (Kulger *et al.*, 2011). Knock-out studies have also shown that *prickle* is essential in mediolateral (M/L) polarization during notochord morphogenesis; *prickle* is expressed in notochord cell membranes, where cells detect interactions with neighboring tissues (Jiang *et al.*, 2005). As *C. intestinalis* contains two *prickle* genes— *prickle1* (pk1) and *prickle2* (pk2)—this study will search for expression of either *prickle* in *M. occulta* and *M. oculata*.

Other notochord-localized genes identified in the *C. intestinalis* transcriptome are *noto6* and *noto17*. These genes are enigmatic in that little is known of their products aside from their presence in urochordate notochords. Studies in the larvacean *Oikopleura dioica*, a member of another class of urochordata, showed diffuse expression of *noto17* in the notochord (Kulger *et al.*, 2011). *Noto6* has been detected in *Molgula tectiformis*, an anural ascidian, (Gyoja *et al.*, 2007) and a homologue has been isolated from human neural tissue, though its function has yet to be described (Hotta *et al.*, 2000). The implications of these results are still under investigation, but the variety of tunicates expressing these genes suggest that they may allow an interesting analysis of phylogeny.

In *C. intestinalis*, *leprecan* is a highly conserved, notochord-localized transcriptional target of Brachyury, a zinc-finger transcription factor that is essential in ascidian notochord development (Di Gregorio *et al.*, 2002). *Ci-leprecan* encodes collagen-producing prolyl-3 hydroxylases and is regulated by *Ci-Bra* via a compact *cis*-regulatory

module (CRM) (Dunn and Di Gregorio, 2009). Dunn and Di Gregorio found that *C. intestinalis* with selectively knocked-out *Ci-leprecan* develop a suboptimal notochord phenotype, which suggests *Ci-leprecan* is essential in notochord development. Another study in *C. intestinalis* found that *Ci-leprecan* is most highly expressed in the notochordal sheath (NS) (Hotta *et al.*, 2000); as the notochord affects specification of surrounding tissue (Liem *et al.*, 2000), this may mean *Ci-leprecan* contributes to tissue specification. While *M. occulta* has been shown to have normal early expression of *Brachyury* in the notochord precursor cells (Takada *et al.*, 2002), Molgolid *leprecan* expression remains unexplored. This study will then build upon past studies by assessing the extent to which the *Ci-Bra1/Ci-leprecan* association is conserved in *M. occulta* and *M. oculata*.

Finally, we will examine the expression of *merlin*, the product of the NF2 tumor suppressor gene. *Merlin* is a linker between proteins embedded in the cell membrane and the actin-cytoskeleton, making it an important contributor to the orchestration of cell-cell interactions (McClatchey and Giovannini, 2005). Probing for *merlin* in *M. oculata* will give insight into the non-autonomous expression patterns at play during the tissue development of ascidian embryos. *Merlin* is also interesting because it is closely linked with nervous tissue, as is evident by how nonfunctional *merlin* correlates with human neurofibromatosis type II, the symptoms of which are schwannomas, ependymomas and meningiomas (Stamenkovic and Yu, 2010).

Materials and Methods

Animal Samples

M. occulta and *M. oculata* specimens were collected on the sand flats of Pointe de Blosson at Station Biologique in Roscoff, France by Dr. Billie Swalla in 1999. Eggs and sperm extracted from the animal gonads were fertilized to raise *M. occulta* and *M. occulta* x *M. oculata* hybrids until embryos hatched. *In situ* samples of 10 life stages (F+1, F+1.5, F+2, F+3, F+4, F+5, F+6, F+7, F+8, F+9) of the *M. occulta*, *M. oculata* and hybrids were fixed in 4% paraformaldehyde and preserved in 70% ethanol; these samples were used in this study. The procedures for insemination, embryo culture and production of hybrids are previously described (Swalla and Jeffery, 1990).

Molgulid transcriptome

Genes selected for analysis were chosen from *M. oculata*, *M. occulta* and *M. occulta* x *M. oculata* hybrid transcriptomes sequenced by Illumina sequencing at Michigan State University. Nucleotide fragments were between 200-250 bp. RNAs were extracted from several different stages of development, sequenced, then reassembled; the lengths of reassembled *prickle*, *noto6*, *noto17*, *leprecan* and *merlin* were 1077 bp, 1154 bp, 960 bp, 888 bp and 1121 bp respectively. Data from the gastrula (F+3), F+4, and F+6 stages were used for analysis.

Sequence Alignments

Protein and nucleotide BLAST was used to confirm the identity of genes selected and to assess protein conservation. All genes were found to be highly conserved in *Molgula tectiformis* and *Ciona intestinalis*. BLASTx was used to derive protein sequences from the *M. occulta*, *M. oculata* and hybrid DNA sequences and to check the orientation of the

DNA sequences. Reverse complements of the transcripts were obtained using Sequencher. All forward nucleotide sequences and protein sequences from the *M. occulta*, *M. oculata* and hybrid transcriptomes were aligned using ClustalW2 (<http://www.ebi.ac.uk/Tools/msa/clustalw2/>). For some genes, there were sections in which one transcriptome did not have a reading; this is being attributed to mistakes inherent in DNA sequencing assembly.

PCR and Subcloning

Primers were designed using DNA and protein alignments. Highly conserved sections at the start and end of each protein sequence were translated using the genetic code. This DNA sequence was then compared with the actual DNA sequences derived by Dr. Titus Brown *et al.* (2011, unpublished data). Each primer is between 20-23 bp and was selected for highly conserved 5' and 3' end sequences and minimal degeneracy. The forward and reverse primers used are listed in Table 1:

Table 1. Forward and Reverse Primer Sequences

Gene	Forward	T _m (°C)	Reverse	T _m (°C)
<i>prickle</i>	5' -GATAGCGGATGTTCC TTGGAAG-3'	55.4	5' -CGCTTCGGATGGGTACAGCTC-3'	60.1
<i>noto6</i>	5' -ATGTATGAAATAACAAGATGG-3'	45.7	5' -TCAGTCAAATAAAGAATATTTAA-3'	43.8
<i>noto17</i>	5' -GAGAGTCAAACATGGATCCG-3'	53.4	5' -TCACTGCGTTTCCCAGGAAGCAT-3'	61.2
<i>leprecan</i>	5' -GAACCTGAAACTGAAGAATTAGC-3'	51.9	5' -GCGTCAGTATCAAGGAACAT-3'	52.4
<i>merlin</i>	5' -AAGAGCATTAAACGTTTGCATTTTC-3'	52.6	5' -CTTCTTTAGCTTGTTTTCAT-3'	49.4

Primer melting points (T_m) ranged from 43.8°C to 61.2°C (see Table 1). PCR conditions were 35 cycles of: 1. Denature at 94°C for 4 minutes, 2. Denature at 94°C for 1 minute, 3.

Anneal at 40°C for 1 minute, 4. Extend at 72°C for 2 minutes. The final PCR cycle was at 72°C for 10 minutes for extension.

PCR inserts were manually extracted from gels and isolated using the GenElute™ Plasmid Miniprep Kit. Following isolation, inserts were incorporated into 3956 bp 4-TOPO® bacterial vectors with T3 and T7 priming sites on either end. These priming sites facilitate linearization for the transcription of sense and anti-sense strands; *Not I* was used to linearize the T3 strand and *Pst I* was used for the T7 strand. On the immediate 6-10 bp following either end of the insert there are also *EcoRI* restriction sites useful for preliminary digests to confirm insert length. Standard transformation procedures were used to ligate plasmids and transform *E. coli* via temperature shock. Subclones were transformed in INVαF' strain of *E. coli* and grown on Kanamycin plates at 37°C. Successfully transformed and proliferated *E. coli* were stored in sterile glycerol and stored at -70°C.

Results

Transcriptome Alignments

BLASTp searches (Altschul *et al.*, 1997) using protein sequences derived from the *M. occulta* and *M. oculata* transcriptomes showed that *prickle* is highly conserved among the ascidians, as well as in more distantly related species like *Homo sapiens* (see Figure 1). The highest BLASTp scores were *Mt-prickle*, *Ci-prickle2* and *Ci-prickle1*. For homologues in other organisms with more than one *prickle* type, *prickle2* was generally the closest match to *Mo-prickle*. Three zinc-binding sites were identified: LIM1 between the 105th and 160th amino acid, LIM2 between the 170th and 220th amino acid and LIM3 between the 230th and 285th amino acid; these are all members of the LIM superfamily, a

group characterized by two highly conserved zinc-finger motifs. A fourth domain, PET_prickle from the Prickle Espinas Testin (PET) superfamily, is found between the 1st and 100th amino acid; this is also characterized by a zinc-finger protein-protein interaction domain. As the molgulid transcriptome sequences are approximately 400 amino acids shorter than that of *M. tectiformis*, it is likely that the full molgulid sequence for prickles has not been isolated (Figure 1). Within the molgulid sequences, there are no significant gaps, meaning that *Mo-prickle* has been highly conserved in the chordates (Figure 1).

Noto6 is well conserved among molgulids but not in more distantly related species like human (Figure 2). The human protein sequence is much longer because a section is absent in all the aligned ascidian sequences (Figure 2); this most likely merely indicates an evolutionary divergence between ascidians and human. Insertions were apparent in the lancelet and human *noto6* homologues approximately halfway into the human protein sequence (Figure 2). No identifiable binding regions were found in BLASTp, though the full sequence was identified as Glyco-tranf-GTA-type superfamily (Marchler-Bauer et al. 2011). Domain hits include the single domain Chondroitin N-acetylgalactosaminyltransferase and multi-domain riboflavin synthase subunit alpha.

A *noto17* BLASTp search yielded one nonspecific hit in the Thioredoxin_like superfamily with a CXXC motif between the 40th and 90th amino acid (Kikuchi et al., 2002). The molgulid *noto17* sequence may be incomplete because the sequence lengths of the transcriptomes are significantly shorter than that of *M. tectiformis*; more rounds of sequencing *Mo-noto17* will help in resolving the full length sequence.

Leprecan was highly conserved among the molgulids, the non-molgulid ascidian *C. intestinalis* and human (Figure 4). A sizable portion of the full sequence appears to be present, but the fact that the transcriptome sequences are shorter than *M. tectiformis* suggests the ends of the sequence may be missing (Figure 4). The molgulid *leprecan* protein sequence had a 20G Fe(II) oxygenase superfamily match between the 110th and 296th amino acid.

Merlin is also highly conserved among the molgulids; there is a low level of conservation with human, but it is worth noting that there were no insertions or deletions present in the human protein sequence. We do not appear to have the full protein isolated, as there are large gaps of information missing between the molgulid transcriptomes themselves and between the transcriptomes and *M. tectiformis* (Figure 5). BLASTp identified an ezrin/radixin/moesin (ERM) protein domain between the first and 210th amino acid; this domain characterizes a phosphatase, which supports the previously established function of *merlin* (McClatchey and Giovannini, 2005). There is a strongly conserved PH-like core between the 210th and 320th amino acid within a FERM-C domain; this site is capable of both peptide and lipid binding (García-Alvarez et al., 2003, Hamada et al., 2000). Upstream there is also a FERM_N and FERM_M domain between the 20th-90th amino acid and 100th-210th amino acid respectively.

PCR and Subcloning

The first PCR reaction had an annealing temperature of 45°C and produced some bands, but none with a proper length. *M. occulta Mo-prickle*, *M. oculata Mo-prickle*, *M. occulta Mo-noto17*, *M. oculata Mo-noto17*, *M. occulta Mo-leprecan*, *M. oculata Mo-leprecan*, *M. occulta Mo-merlin*, and *M. oculata Mo-merlin* were amplified by PCR with

an annealing temperature of 40°C (Figure 7). PCR was repeated for *Mo-noto6* at 38°C, but this temperature also failed to amplify *Mo-noto6* (Figure 8). The failure of my *Mo-noto6* primers may be attributed to the apparent incompleteness of the *Mo-noto6* sequence; further sequencing may reveal more inclusive primer sites.

Subclones were successfully grown for *Mo-prickle*, *Mo-noto17*, *Mo-leprecan* and *Mo-merlin* for both *M. occulta* and *M. oculata*. When 100 µl of *E. coli* transformed with these genes were grown on Kanamycin plates, *Mo-prickle* for *M. oculata* produced the most colonies (31) and *Mo-leprecan* for *M. oculata* produced the least colonies (2). Minipreps digested with *EcoRI* revealed that the inserts for *Mo-noto17*, *Mo-leprecan* and *Mo-merlin* were shorter than the anticipated lengths of 960 bp, 888 bp, and 1121 bp respectively (Figure 9). *Mo-prickle* for *M. oculata* consistently appeared at approximately 1077 bp (Figure 10), as anticipated from the *Mo-prickle* sequence. *Mo-prickle* for *M. occulta*, however, appeared notably shorter than *Mo-prickle* for *M. oculata* (Figure 10).

RNA probes were successfully made for the T7 strand of *Mo-prickle* (Figure 11), but not for the T3 strand; a low concentration of DNA in the T3 transcription reaction may account for the insignificant yield of T3 probes.

Discussion

Prickle in the Molgulids

The *M. occulta* and *M. oculata* *prickle* sequences aligned in this study may imply the presence of more than one *prickle* in the molgulids (Figure 1). *Prickle* is well conserved between *M. occulta* and *M. oculata*, but there are several permutations in the aligned amino acid sequences that may justify a distinction between *prickle* in these species. The relative sizes of *prickle* in *M. occulta* and *M. oculata* appear to be different

in the DNA segments seen in the miniprep *EcoRI* digestion (Figure 10). This could potentially imply a deletion in the *prickle* sequence of *M. occulta*, but only sequencing the isolated genes may give immediate conclusive results. A search for restriction sites within *prickle* using Sequencher found that *prickle* in *M. occulta* and *M. oculata* have *Hind III* and *Xba I* restriction sites in common, but otherwise have several unshared restriction sites; *prickle* in *M. occulta* has *Kpn I*, *BamHI*, and *Pst I* restriction sites (Figure 12) while *prickle* in *M. oculata* has *Sac I* and *Bst XI* restriction sites (Figure 13). The difference in *prickle* sequences causing these different restriction sites may also support the defining of *Mo-prickle1* and *Mo-prickle2*. The presence of *Ci-prickle1* and *Ci-prickle2* in the ascidian *Ciona intestinalis* (Hotta *et al.*, 2000) strengthens the feasibility of more than one *prickle* being present in molgulid ascidians.

Molecular nature of Molgulid genes

The primers designed to isolate *prickle*, *noto6*, *noto17*, *leprecan* and *merlin* for *M. occulta* and *M. oculata* required exceptionally low annealing temperatures. Many A-T base-pairs translate to lower annealing temperatures because A-T hydrogen bonding is less efficient than C-G hydrogen bonding at overcoming the initial inertia of free-moving primers. It is then possible that these low annealing temperatures are due to the ubiquity of A-T bonds in the gene sequences. High A-T content in the DNA sequences of *prickle*, *noto6*, *noto17*, *leprecan* and *merlin* in *M. occulta* and *M. oculata* is readily noticeable at the ends of the genes, where primers were designed. It is possible that the ascidians are A-T rich due to environmental pressures in the same way thermophiles are C-G rich (Basak *et al.*, 2010). A-T ubiquity in this sense may then be a consequence of being native to benthic, low-temperature environments.

Future Studies

The immediate next step in this project is the sequencing of the genes isolated during this study. This will confirm their identity and allow the project to move forward into an investigation of gene expression via *in-situ* hybridization. The genes I focused on do not contain the restriction sites used to linearize the T3 (*Not I*) and T7 (*Pst I*) strands for probe synthesis, with the exception of *prickle* in *M. occulta*. This *prickle* appears to have a downstream *Pst I* restriction site (Figure 12). Future researchers may resolve this conflict by choosing a different bacterial vector, in which, the restriction sites used to linearize sense and anti-sense strands are not found in the gene.

Once probes are synthesized for *Mo-prickle*, *Mo-noto17*, *Mo-leprecan* and *Mo-merlin*, *in-situ* hybridizations may be done in the neurula and tail-bud stage embryos of *M. occulta*, *M. oculata* and *M. occulta* x *M. oculata* hybrids. Dr. Billie Swalla has preserved neurula and tail-bud stage embryos of these organisms available for such a study; animals were collected in Roscoff, France in 1999. The *in-situ* results for *M. oculata* are expected to resemble those found in the ascidian *Ciona intestinalis* (Hotta *et al.*, 2000); in the neurula stage, the probes will appear down the A/P axis of the embryo, while in the tail-bud stage the probes will mainly illuminate the notochord. An *M. occulta* x *M. oculata* hybrid *in-situ* is also expected to show an illuminated notochord; though there are fewer notochord cells present (Swalla and Jeffery, 1996), these hybrids have nonetheless completed convergence and elongation and thus produce the molecular transcripts required to orchestrate this event.

Results of the *M. occulta in-situ* are more elusive. No expression of these genes in the tail-bud stage of *M. occulta* would support the hypothesis that these genes are

required for the full development of a notochord. However, if these genes *are* expressed, it may mean they are not necessarily a requirement for late-stage notochord development. Knock-out studies have shown that without *prickle*, proper development in ascidians does not occur (Jiang *et al.*, 2005); suboptimal development has also been described in *leprecan* knock-out experiments (Dunn and Di Gregorio, 2009). If this holds true for *noto6*, *noto17* and *merlin*, it would mean these genes are a *requirement* for notochord development, but not necessarily the key genes involved in the converge-and-extend event absent in *M. occulta*.

This second scenario immediately raises a question of efficiency: why produce a transcriptional product that doesn't lead to a morphological change? It is possible that expression of these genes in late stages of anural embryonic development may be vestigial signals; natural selection does not necessarily select for complete elimination of gene expression if the expression does not significantly impede the organism.

Another important question to consider while studying molgulid tail-loss is what kind of factors facilitated tail-loss. These factors may be active or passive: Active selective factors mean taillessness was *selected for* by pressures in the environment, while passive factors mean tail-loss simply was not *selected against*. Passive selection would be made possible by the genetic plasticity observed in molgulids (Iannelli *et al.*, 2007).

In-situ hybridization is the ultimate end of this particular study, but it is not the only or even the most conclusive way to investigate the overarching question of this project: are *prickle*, *noto6*, *noto17*, *leprecan* and *merlin* required for the convergence and extension of notochordal cells? Aside from *in-situs*, a knock-out gene experiment for

each of these genes in anural and urodele molgulids would be compelling as a way to directly determine the importance of each gene in the converge-and-extend event.

Acknowledgements

I would like to thank BEACON, National Science Foundation and the American Society for Cell Biology for their financial support, as well as everyone at Friday Harbor Laboratories who has made my research possible. I would especially like to thank my mentor, Dr. Billie J. Swalla, Dr. Sophie George, Peter X. Wu, Paul Hausch, Joie Cannon, Dr. Kenneth Halanych, Dr. Brad Schuster, Silvia Sipulveda, Dr. Titus Brown, Elijah Lowe and Max Maliska for all of their input and guidance.

Literature Cited

- Altschul S.F., Madden T.L., Schäffer A.A., Zhang J., Zhang Z., Miller W., Lipman D.J. (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* 25:3389-3402.
- Basak S., Mukhopadhyay P., Gupta S., Ghosh T. (2010) Genomic adaptation of prokaryotic organisms at high temperature. *Bioinformatics* 4 (8): 352-256
- Dehal P., Satou Y., Campbell R. K., Chapman J., Degnan B., De Tomaso A., Davidson B., Di Gregorio A., Gelpke M., Goodstein D.M., Harafuji N., Hastings K.E., Ho I., Hotta K., Huang W., Kawashima T., Lemaire P., Martinez D., Meinertzhagen I.A., Necula S., Nonaka M., Putnam N., Rash S., Saiga H. Satake M., Terry A., Yamada L., Wang H.G., Awazu S., Azumi K., Boore J., Branno M., Chin-Bow S., DeSantis R., Doyle S., Francino P., Keys D.N., Haga S., Hayashi H., Hino K., Imai K.S., Inaba K., Kano S., Kobayashi K., Kobayashi M., Lee B.I., Makabe K.W., Manohar C., Matassi G., Medina M., Mochizuki Y., Mount S., Morishita T., Miura S., Nakayama A., Nishizaka S., Nomomoto H., Ohta F., Oishi K., Rigoutsos I., Sano M., Sasaki A., Sasakura Y., Shoguchi E., Shin-i T., Spagnuolo A., Stainier D., Suzuki M.M., Tassy O., Takatori N., Tokuoka M., Yagi K., Yoshizaki F., Wada S., Zhang C., Hyatt P.D., Larimer F., Detter C., Doggett N., Glavina T., Hawkins T., Richardson P., Lucas S., Kohara Y., Levine M., Satoh N., and Rokhsar D.S. (2002) The draft genome of *Ciona intestinalis*: Insights into chordate and vertebrate origins. *Science* 298: 2157-2167
- Di Gregorio A., Harland R.M., Levine M., Casey E.S. (2002) Tail morphogenesis in the ascidian, *Ciona intestinalis*, requires cooperation between notochord and muscle. *Developmental Biology* 244 (2): 385–395
- Dunn M.P. and Di Gregorio A. (2009) The evolutionarily conserved leprecan gene: its regulation by Brachyury and its role in the developing *Ciona* notochord. *Developmental Biology* 328 (2): 561-574
- García-Alvarez B., de Pereda J.M., Calderwood D.A., Ulmer T.S., Critchley D., Campbell I.D., Ginsberg M.H., Liddington R.C. (2003) Structural determinants of integrin recognition by talin. *Molecular Cell* 11(1): 49-58
- Gyoja F., Satou Y., Shin-i T., Kohara Y., Swalla B.J., Satoh N. (2007) Analysis of large scale expression sequenced tags (ESTs) from the anural ascidian, *Molgula tectiformis*. *Developmental Biology* 307: 460-482
- Hamada K., Shimizu T., Matsui T., Tsukita S., Hakoshima T. (2000) Structural basis of the membrane-targeting and unmasking mechanisms of the radixin FERM domain. *European Molecular Biology Organization, Japan* 19(17): 4449-62

- Hotta K., Takahashi H., Asakura T., Saitoh B., Takatori N., Satou Y., Satoh N. (2000) Characterization of Brachyury-Downstream Notochord Genes in the *Ciona intestinalis* Embryo. *Developmental Biology* 224: 69-80
- Huber J.L., Burke da Silva K., Bates W.R., Swalla B.J. (2000) The evolution of anural larvae in molgulid ascidians. *Cell & Developmental Biology* 11 (6): 419-426
- Iannelli F., Griggio F., Pesole G., Gissi C. (2007) The mitochondrial genome of *Phallusia mammillata* and *Phallusia fumigata* (Tunicata, Ascidiacea): high genome plasticity at intra-genus level. *BMC Evolutionary Biology* 7 (155). (<http://www.biomedcentral.com/1471-2148/7/155>)
- Jiang D., Munro E.M., Smith W.C. (2005) Ascidian prickle regulates both mediolateral and anterior-posterior cell polarity of notochord cells. *Current Biology* 15 (1): 79-85
- Kikuchi M., Doi E., Tsujimoto I., Horibe T., Tsujimoto Y. Functional analysis of human P5, a protein disulfide isomerase homolog. (2002) *J Biochem* 132(3): 451-5
- Kulger J.E., Pierre K., Bouquet J., Jiang D., Di Gregorio A. (2011) Evolutionary changes in the notochord genetic toolkit: a comparative analysis of notochord genes in the ascidian *Ciona* and the larvacean *Oikopleura*. *BMC Evolutionary Biology* 11: 21
- Liem K.F. Jr, Jessell T.M., Briscoe J. (2000) Regulation of the neural patterning activity of sonic hedgehog by secreted BMP inhibitors expressed by notochord and somites. *Development* 127 (22): 4855-66
- Marchler-Bauer A., Lu S., Anderson J.B., Chitsaz F., Derbyshire M.K., DeWeese-Scott C., Fong J.H., Geer L.Y., Geer R.C., Gonzales N.R., Gwadz M., Hurwitz D.I., Jackson J.D., Ke Z., Lanczycki C.J., Lu F., Marchler G.H., Mullokandov M., Omelchenko M.V., Robertson C.L., Song J.S., Thanki N., Yamashita R.A., Zhang D., Zhang N., Zheng C., Bryant S.H. CDD: a Conserved Domain Database for the functional annotation of proteins. (2011) *Nucleic Acids Research (Database Issue)*: D225-9
- McClatchey A.I. and Giovannini M. (2005) Membrane organization and tumorigenesis—the NF2 tumor suppressor, Merlin. *Genes & Development* 19 (19): 2265-2277
- Milligan M. (1946) Trichrome stain for formalin-fixed tissue. *American Journal of Clinical Pathology, Technical Section* 10: 184-85
- Scholz C.B. and Technau U. (2003) The ancestral role of Brachyury: expression of *NemBra1* in the basal cnidarian *Nematostella vectensis* (Anthozoa). *Development and Genes Evolution* 212 (12): 563-570
- Stamenkovic I. and Yu Q. (2010) Merlin, a "Magic" Linker Between the Extracellular Cues and Intracellular Signaling Pathways that Regulate Cell Motility, Proliferation, and Survival. *Current Protein & Peptide Science* 11 (6): 471-484.

Swalla B.J. and Jeffery W.R. (1990) Interspecific hybridization between an anural and urodele ascidian: Differential expression of urodele features suggests multiple mechanisms control anural development. *Developmental Biology* 142: 319-334

Swalla B.J., Makabe K.W., Satoh N., Jeffery W.R. (1993) Novel genes expressed differentially in ascidians with alternate modes of development. *Development* 119: 307-318

Swalla B.J. and Jeffery W.R. (1996) Requirement of the *Manx* gene for expression of chordate features in a tailless ascidian larva. *Science* 274 (5290): 1205-8

Swalla B.J. (2004) Protochordate Gastrulation: Lancelets and Ascidians, in *Gastrulation: From Cells to Embryo*, Chapter 10 (Stern CD, ed) p.139-149. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY

Takada N., York J., Davis J.M., Schumpert B., Yasuo H., Satoh N., Swalla B.J. (2002) Brachyury expression in tailless Molgulid ascidian embryos. *Evolution & Development* 4 (3): 205-211

Tao H, Suzuki M, Kiyonari H., Abe T., Sasaoka T., Ueno N. (2009) Mouse *prickle1*, the homolog of a PCP gene, is essential for epiblast apical-basal polarity. *PNAS* 106 (34): 14426-14431

FIGURE LEGENDS

Figure 1. Protein alignment for *prickle* including *M. occulta* and *M. oculata* transcriptome sequences and sequences from another anural molgulid (*M. tectiformis*), a urodele non-molgulid ascidian (*Ciona intestinalis*) and human. Yellow indicates a LIM1 domain, blue indicates a LIM2 domain, green indicates a LIM3 domain and fuchsia indicates a PET_prickle domain. Amino acids conserved between all species are indicated by an asterisk (*), amino acids with the same charge are indicated by a period (.) and significant but imperfect matches are indicated by a semicolon (:).

Figure 2. Protein alignment for *noto6* including *M. occulta* and *M. oculata* transcriptome sequences and sequences from another anural molgulid (*M. tectiformis*), a urodele non-molgulid ascidian (*C. intestinalis*), lancelet and human. Amino acids conserved between all species are indicated by an asterisk (*), amino acids with the same charge are indicated by a period (.) and significant but imperfect matches are indicated by a semicolon (:).

Figure 3. Protein alignment for *noto17* including *M. occulta* and *M. oculata* transcriptome sequences and sequences from another anural molgulid (*M. tectiformis*) and two urodele non-molgulid ascidians (*C. intestinalis* and *Oikopleura dioica*). Yellow indicates a Thioredoxin_like domain. Amino acids conserved between all species are indicated by an asterisk (*), amino acids with the same charge are indicated by a period (.) and significant but imperfect matches are indicated by a semicolon (:).

Figure 4. Protein sequence alignment for *leprecan* including hybrids of *M. occulta* and *M. oculata*. In addition there are sequences for *M. tectiformis*, *C. intestinalis* and human. Yellow indicates a 20G Fe(II) oxygenase domain. Amino acids conserved between all species are indicated by an asterisk (*), amino acids with the same charge are indicated by a period (.) and significant but imperfect matches are indicated by a semicolon (:).

Figure 5. Protein sequence alignment for *merlin* including *M. occulta* and *M. oculata*, as well as *M. tectiformis*, *C. intestinalis*, an acorn worm (*Saccoglossus kowalevskii*) and human. Yellow indicates an ezrin/radixin/moesin (ERM) domain. White diamonds (◇) indicates a FERM_N domain, black dots (●) indicate a FERM_M domain and white pentagons (△) indicate a FERM_C domain with a PH-like core. Amino acids conserved between all species are indicated by an asterisk (*), amino acids with the same charge are indicated by a period (.) and significant but imperfect matches are indicated by a semicolon (:).

Figure 6. Results of PCR with 45°C annealing temperature. Lanes starting from the far left are: 1 kb ladder, *prickle* for *M. occulta*, *prickle* for *M. oculata*, *noto17* for *M. occulta*, *noto17* for *M. oculata*, *noto6* for *M. occulta*, *noto6* for *M. oculata*, *leprecan* for *M. occulta*, *leprecan* for *M. oculata*, *merlin* for *M. occulta* and *merlin* for *M. oculata*. Expected bands for *prickle*, *noto17*, *noto6*, *leprecan* and *merlin* are 1077 bp, 960 bp, 1154 bp, 888 bp and 1121 bp respectively; the bands that appear are too small to be correct.

Figure 7. Results of PCR with 40°C annealing temperature. Lanes starting from the far left are: 1 kb ladder, *prickle* for *M. occulta*, *prickle* for *M. oculata*, *noto6* for *M. occulta*, *noto6* for *M. oculata*, *noto17* for *M. occulta*, *noto17* for *M. oculata*, *leprecan* for *M. occulta*, *leprecan* for *M. oculata*, *merlin* for *M. occulta* and *merlin* for *M. oculata*. Expected bands for *prickle*, *noto17*, *noto6*, *leprecan* and *merlin* are 1077 bp, 960 bp, 1154 bp, 888 bp and 1121 bp respectively.

Figure 8. Results of PCR with 38°C annealing temperature. Lanes starting from the far left are: 1 kb ladder, *noto6* for *M. occulta*, *noto6* for *M. oculata*, uncut *merlin* for *M. oculata* (miniprep DNA) and *EcoRI* digest of *merlin* for *M. oculata*.

Figure 9. Results of first miniprep. Lanes starting from the far left of Gel 1 are: 1 kb ladder, uncut *noto17* for *M. occulta* culture 1, *EcoRI* digest of *noto17* for *M. occulta* culture 1, uncut *noto17* for *M. occulta* culture 2, *EcoRI* digest of *noto17* for *M. occulta* culture 2, uncut *noto17* for *M. oculata* culture 3, *EcoRI* digest of *noto17* for *M. oculata* culture 3, uncut *noto17* for *M. oculata* culture 4, *EcoRI* digest of *noto17* for *M. oculata* culture 4, uncut *noto17* for *M. oculata* culture 5, *EcoRI* digest of *noto17* for *M. oculata* culture 5, uncut *merlin* for *M. occulta* culture 6, *EcoRI* digest of *merlin* for *M. occulta* culture 6, uncut *merlin* for *M. occulta* culture 7 and *EcoRI* digest of *merlin* for *M. occulta* culture 7.

Lanes starting from the far left of Gel 2 are: 1 kb ladder, uncut *merlin* for *M. occulta* culture 8, *EcoRI* digest of *merlin* for *M. occulta* culture 8, uncut *merlin* for *M. oculata* culture 9, *EcoRI* digest of *merlin* for *M. oculata* culture 9, uncut *merlin* for *M. oculata* culture 10, *EcoRI* digest of *merlin* for *M. oculata* culture 10, uncut *merlin* for *M. oculata* culture 11, *EcoRI* digest of *merlin* for *M. oculata* culture 11, uncut *prickle* for *M. oculata* culture 12 and *EcoRI* digest of *prickle* for *M. oculata* culture 12.

Figure 10. Results of second miniprep. Lanes starting from the far left of Gel 1 are: 1 kb ladder, uncut *prickle* for *M. oculata* culture 1, *EcoRI* digest of *prickle* for *M. oculata* culture 1, uncut *prickle* for *M. oculata* culture 2, *EcoRI* digest of *prickle* for *M. oculata* culture 2, uncut *prickle* for *M. oculata* culture 3, *EcoRI* digest of *prickle* for *M. oculata* culture 3, uncut *prickle* for *M. oculata* culture 4, *EcoRI* digest of *prickle* for *M. oculata* culture 4, uncut *prickle* for *M. oculata* culture 5, *EcoRI* digest of *prickle* for *M. oculata* culture 5, uncut *prickle* for *M. oculata* culture 6, *EcoRI* digest of *prickle* for *M. oculata* culture 6, uncut *prickle* for *M. occulta* culture 7 and *EcoRI* digest of *prickle* for *M. occulta* culture 7.

Lanes starting from the far left of Gel 2 are: 1 kb ladder, uncut *prickle* for *M. occulta* culture 8, *EcoRI* digest of *prickle* for *M. occulta* culture 8, uncut *prickle* for *M. occulta* culture 9, *EcoRI* digest of *prickle* for *M. occulta* culture 9, uncut *leprecan* for *M. occulta* culture 10, *EcoRI* digest of *leprecan* for *M. occulta* culture 10, uncut *leprecan* for *M. occulta* culture 11, *EcoRI* digest of *leprecan* for *M. occulta* culture 11, uncut *leprecan* for *M. occulta* culture 12 and *EcoRI* digest of *leprecan* for *M. occulta* culture 12.

Figure 11. Dot blot test for T3 and T7 RNA probes. Y axis shows 1/10, 1/50, 1/100 and 1/500 dilutions for the probes, while the X axis shows, from left to right, the control probe, T3 probe and T7 probe. A purple dot signifies a successful probe.

Figure 12. Sequencher representation of restriction sites in *Mo-prickle* from *Molgula occulta*.

Figure 13. Sequencher representation of restriction sites in *Mo-prickle* from *Molgula oculata*.

Figure 1

```

Molgula.oculata.prickle -----
Molgula.tectiformis.prickle MLKFQHKNALHTLVSCMTGRPCTKCDPGICPGFALHEWRKVAHCKCKGVD 50
Molgula.occulata.prickle -----
Ciona.intestinalis.prickle -----MTMPAAATEQTRGTMPSNIDPKSAGLDQDIVIR----- 33
Human.prickle -----MVTVMPLMEKTI SKLMFDFQRNST----- 25

Molgula.oculata.prickle -----
Molgula.tectiformis.prickle YHVGDDTNQTSENNVSVDLPAISNKFEANDIMVGHGHNATPNNNNIDHYDR 100
Molgula.occulata.prickle -----
Ciona.intestinalis.prickle ---GPTENRVRRRRQSRRQASVR-----HN-----R 55
Human.prickle -----

Molgula.oculata.prickle -----SDDSGCSLEDCAWTPPGLTPKQVHNYF SKLPEDRIF 37
Molgula.tectiformis.prickle NQRSHKTGTRGLFNDDTDSGCVLEECAWVPPGLSPKQAQAYF SKLPEDRIF 150
Molgula.occulata.prickle -----DDSGCSLEECAWVPPGLTPKQVHNYF SKLPEDRIF 36
Ciona.intestinalis.prickle NSASDEE-----NDGSGCALEEYAWVFPNLT PDQVRYF TSLPEDKVF 99
Human.prickle -----SDDSGCALEEYAWVPPGLKPEQVHQYYSC LPEEKVF 62
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle FTDSIGEKHRI RQLLQQLPPHDNEVRYCNDLSEEEKHELKIFSEQRKMEA 87
Molgula.tectiformis.prickle FTDSIGEKHRI RQLLQQLPPHDNEVRYCNDLSEEEKHELKIFSEQRKTEA 200
Molgula.occulata.prickle FTDSIGEKHRI RQLLQQLPPHDNEVRYCNDLSEEEKHELKIFSEQRKMEA 86
Ciona.intestinalis.prickle LVDSIGDKYRV RQLLQQLPPHDNKVCYCNLSDDEEKRELRLFSEQRKDY 149
Human.prickle VVNSPGEKLR I KQLLHQLPPHDNEVRYCNSLDEEEKRELKLFSSQRKREY 112
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle LGRGTARPPPSNI PSAICENCGMRIEGGDI AVFASRAGRGVCWH PACFVC 137
Molgula.tectiformis.prickle LGRGTARPPPNIP PAICENCGYH INGGDI AVFASRAGCAVCWH PNCFVC 250
Molgula.occulata.prickle LGRGTARPPPSNI PSAICENCGMRIEGGDI AVFASRAGRGVCWH PACFVC 136
Ciona.intestinalis.prickle LGCGKIR I LPLNTPGTPCSECGILVKGGDI AVASRAEFP GCMWH PACFVC 199
Human.prickle LGRGNVRPFPVMTGAI CQCQGGQ INGGDI AVFASRAGHGV CWH PPFVC 162
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle SVCDELLVDLIYFHQDGLYCGRRHHAETLKPRC SACDEI I FADECTEAE 187
Molgula.tectiformis.prickle SVCDELLVDLIYFHQDGLYCGRRHHAETLKPRC SACDEI I FADECTEAE 300
Molgula.occulata.prickle SVCDELLVDLIYFHQDGLYCGRRHHAETLKPRC SACDEI I FADECTEAE 186
Ciona.intestinalis.prickle SVCRELLVDLIYFYQDGRLYCGRRHHAETLKPRC SACDEI I FSDECTEAE 249
Human.prickle TVCNELLVDLIYFYQDGKIYCGRRHHAETLKPRC AACDEI I FADECTEAE 212
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle RHWHMNHFCFCFCDVVLGGQRYIMRDGKPYCTGCFEQHYAEYCDTCGEVI 237
Molgula.tectiformis.prickle RHWHMNHFCFCFCEVVLGGQRYIMRDGKPYCTS CFQTYAEYCDTCGDI 350
Molgula.occulata.prickle RHWHMNHFCFCFCDVVLGGQRYIMRDGKPYCTGCFEQLYAEYCDTCGEVI 236
Ciona.intestinalis.prickle RHWHMDHFCFCFCDVVLGGQRYIMRDGKPNCTQC FEALYAEYCDMCGDLI 299
Human.prickle RHWHMNHFCFCFCETVLGGQRYIMKEGRPYCCHCFESLYAEYCDTCAQHI 262
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle GLDAGQM QYEGQHWHATDGCFCARSKKSL LQRPF LPKHGQIFCSKA SQ 287
Molgula.tectiformis.prickle GLDAGQM QYEGQHWHATDRCFSCARCKKSL LERPF LPKHGQIFCSKA SH 400
Molgula.occulata.prickle GLDAGQM QYEGQHWHATDGCFCARCKKSL LQRPF LPKHGQIFCSKA SQ 286
Ciona.intestinalis.prickle GLDAGQM QYEGQHWHATDNCFCNRCRKS LLGRPF LPKHGRIFCSKA SQ 349
Human.prickle GIDQGOMTYDGOHWHATE TCFCCAHCCKSL LGRPF LPKGQIFCSRAQ SA 312
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle NEDQIHSESDS-----QYENPSVTISHNVRSLNLENLSLHEKANWEN-E 331
Molgula.tectiformis.prickle GEDQLHSESDS-----QYEKATTPVSHNVRSLNLENLSLHEKN-WDNSS 444
Molgula.occulata.prickle NEDQIHSESDS-----QYENPSVPISHNVRSLNLENLSLHEKANWEN-E 330
Ciona.intestinalis.prickle GEDPHGSESDSQHSSSQYENPQLPTSHNVRSLNLDNLSI HDKP-WEDK 398
Human.prickle GEDPNGSDSD-----SAFQNAKAKESR--RSAKIGKNKGKTEEPMLNQHS 356
* * * * * * : * * * * * * . * * * * * : * * * * * *

Molgula.oculata.prickle LCDKVHD--LPVDLSELYPSEAIVA----- 354
Molgula.tectiformis.prickle SVEKSQSDSLPVDLNDLYPSDAIVAS--QHNKCLR LAKNGRID----- 485
Molgula.occulata.prickle LCDKVHD--LPVDLSELYPSEAIVA----- 353
Ciona.intestinalis.prickle ELSPASN-NVFIDAADMYPTSAAVAAS TRYSGHTRPSPYLDGMDPVNA 447
Human.prickle QLQVSSN-RLSADVDPLSLQMDMLSSSQTPSLNRDPIWRSRE----- 398
. : : * : : :

Molgula.oculata.prickle -----
Molgula.tectiformis.prickle -----DYERTKNTKNVASESAAQP---VAS-----FPQNTYNS 515
Molgula.occulata.prickle -----
Ciona.intestinalis.prickle EMVTENDAGFKGAATSRKT VTDVTSPTSTVSSRRTTSKNGVQFPQNTYNS 497
Human.prickle -----EPYHYGNKMEQNQTQSP-----LQLLSQC 422

Molgula.oculata.prickle -----

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Molgula.tectiformis.prickle	TDSSGYNSSSTIDAMDHKSRGRNGQTKVVPSTLNSTCSTA-SQATTCSA	564
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	TDSSGYNSSSTLDAIEH-----QQNAALKAAMGSNYSGKSKQTPCSK	540
Human.prickle	NIRTSYSPGG-----QGAGAQPPEMWGKHFNSNPKRSSSLAMT	458
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	MSNSDKAFPCMGHVPASEFVPPYRVNCSESNKNNNIVQKRIESILSSTQD-	613
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	RFQNGE---DGHVSATEFTPFHPAAPRASPTIIGSRKLAPEIKKTIDS	586
Human.prickle	-----GHAGS----FIKECREDYYPGRILRSQESYSMDSSQSFS	492
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	TQNSSVWQTRRPSVEKIS---AFKSVSDSNLKHPIINGPIAPRAKYP	660
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	LTKATEIDNKSPVNVAS---MLPKSAVPIAPARARYAPSLTPSPSTAA	633
Human.prickle	ETRGSIQVPKYEEEEEEGGLSTQQCRTRHPISSLKYTEDMTPTQTPRG	542
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	SKQSD-WSDANQSLSKNEPARR-----VSDITPPSNYFKDTP-----	696
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	SELTSPWMHKSHARTDSPDREFPSPVPVRSPTESKEHSSPLQRSVS	683
Human.prickle	SMESLALSNTGLSADGGAKRQE---HLSRFMPDLKSDSGMN---VS	584
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	-RGATRRLSENNIGL-----RLNT----TSSTGSNNSQPRGILKK	731
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	ERLANKRRSREPISLPEQTISEHPRLRSDDKHVSVDKTSPELKSILKK	733
Human.prickle	EKLSNMGTLNSSM-----QFRSAESVRSLLSAQQYQEMEGNLHQ	623
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	SRSQV-----ESGNIS-----DLHTPTDETPVSP-----IF--PEP	760
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	SRNPSKSFNRERGLSGSLDRLEEFHRKSDVMKYASDDEGAGFGDAQG	783
Human.prickle	LSNPIGYRDLQSHGRMHQS---FDFDGGMAGSKLPGQE---GVRIQPM	665
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	NTPPFNRAGRLNQSARFP-----NAPGSPESKSTTNYFS-----	794
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	DFSSFQRGQRLYSSARFPPEEVTEKPRSQNGGRPRSQHRTRFKDNSALDR	833
Human.prickle	SERTRRRRATSRDDNRRFR-----PHRRRRRRSRSDNALHLASER----	705
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	-----ECEKK---TCSKKLRRTKSTDFTSK-	816
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	THSALNLDELCAIARRNPKPGKTC SKLSGKSTCSKKLKRTRSTDFAFER	883
Human.prickle	-----EAI SRLKDRPPLRAREDYQFMRQRSFQESMGHSRRDLYGQC	748
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	----ATGASKKRQARFANDVPDEHDSWCSTCTSSSDSDYERWDKFDN	862
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	SAATPTSSRKNRRTKRFVED--EEDGWCSTCTSSNDSDYERWDGLGTS	931
Human.prickle	PRTVSDLALQNAFGDRWGPY--FAEYDWCSTCSSSE--SDNE---GYFLG	792
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	VSMTSSPHHQRRST----EFNLTHLQNLQ-----QQAKLRYGVQST-	899
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	PPTSPLSAMRRGSAVGVVNMTRQPPHPLANADSALAASAAGFNSNG	981
Human.prickle	EPIQPAPARLR-----YVTSDELLH-----KYSSYGLPKS-	821
Molgula.oculata.prickle	-----	
Molgula.tectiformis.prickle	---SALPK---YHHSRHSRRHHKNCVIM	922
Molgula.occulta.prickle	-----	
Ciona.intestinalis.prickle	VYRPSMPRNFSTTSHMRYRRRQKKHCIVM	1011
Human.prickle	---STLGG---RQLHSRKRQKSKNCIIS	844

Figure 2

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Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 MAVRSRRPWMSVALGLVLGFTAAASWLIAPRVAELSERKRRGSSSLCSYGR 50

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 SAAGPRAGAQQPLFPQQRPRQEQSPPPARQDLQGPPLPEAAPGITSFRS 100

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 SPWQQPPPLQQRRRGREPEGATGLPGAPAAEAGEPEEEDGGAAGQRDRGRP 150

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 GSSHNGSGDGGAAAPSARPRDFPYVGVMTAQKYLGSRALAAQRTRARFIP 200

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 GHVEFFSSQQPPNAGQPPPLPVIALPGVDDSYPPQKKSFMMIKYMHDHY 250

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 LDKYEFMRADDDVYIKEDKLEEFRLRSLNSSKPLYLQGTGLGNIEELGKL 300

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 GLEPGENFCMGGPGMIFSRVLRMRVPHIGECLREMYTHEDVEVGRCVR 350

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 -----MRFLRGLNILRIMTGIMLGFLSRIA 25
-----MFGRSLLRRPGFPLVPVCAGIVVGYILRGL 29
RFGGTQCWVSYEMQQLFHENYEHNRKGYIQDLHNSKIHAATLHPNKRPA 400

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 FEQ---LRGSTRLTVGTHMDNIKSLDCPKCPIDDHCTLCEPTHVKNPTEN 72
FDS---TFLS--IIEEKRYKKMSSEKESTPAHCKPCPLCALCSDCWRHDA 74
YQYRLHNYMLSRKISELRYRTIQLHRESALMSKLSNTEVSKEDQQLGVIP 450

Molgula.oculata.noto6 -----
Molgula.occulata.noto6 -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6 -----
Lancelet.noto6 -----
Human.noto6 -----MYEITRWYYFDDNYLYDIINEKP-KVPLIGQWKEETQAVTQ 40
-----EETQAVTQ 8
-----MYEITRWYYFDDNYLYDIINDEP-KVLLIGQWAEIEITAVTQ 40
IPSAKPKGMYEVTRWLQFDEHYVYDIINEEP-KIPLVGHWLEEVRAVTH 121
IEEPYRPKTMYEIVRFHFTEKHLIENLDDDP-RIGLIGRWKEDVQDIQE 123
SNFHQPFRERNEVIEWEFLTGKLLYSAENQPPRQSLSSILRTALDDTVL 500

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Molgula.oculata.noto6      IALEYINI-NKMMGRTWKLLKRENGYIQSDVKHGTEYMIDLKVKSMQDDV 89
Molgula.occulata.noto6    IALEYINI-NKMMGRTWKLLKRENGYIQSDVKHGTEYVIDLKVKSMQDDV 57
Molgula.tectiformis.noto6 VALEYINT-NEILGRTWKIKRENGYLSKSDVFQGREYIFDIKAESMQDGI 89
Ciona.intestinalis.noto6 VAMDYINA-NRMLGQSSLSVLKLDHGYLRRDALRGTEYILDVVVQPASHNF 170
Lancelet.noto6            QAVRIISK-EEKT--QWRLAKLNYGYKRIDAFRGMDEVLDMEVAKADGTE 170
Human.noto6              QVMEMINENAKSRGLIDFKEIQYGYRRVNFPMHGVEYILDLLLLYKRHKG 550
      .: * . . : : . ** : : : * :*:*:

Molgula.oculata.noto6      -----
Molgula.occulata.noto6    -----
Molgula.tectiformis.noto6 -----
Ciona.intestinalis.noto6  S----- 171
Lancelet.noto6            KGRKLVHLVRSTRPHVRLMSLQNVAAQQTVVHFIVPLSRVTDRLRDFMKVY 220
Human.noto6              RKLTPVRRHAYLQQLFSPKPFRETEELDVN----- 581

Molgula.oculata.noto6      -----EETFRTTVVHPL--QSSYEVQSFI 111
Molgula.occulata.noto6    -----EETFRTTIVHPL--ESTYEVESFI 79
Molgula.tectiformis.noto6 -----VRSFRIQLLHPFNHQNEVEVVSIV 113
Ciona.intestinalis.noto6  -----YFINERMFRVNLVHPLKSMGEAEVVRVQ 199
Lancelet.noto6            EKVCLETGAAVSLGLVLFHSFIQSFTHSFTPSFLHSFT-HSPLPTCRCM 269
Human.noto6              -----SLVESINSETQSFISNSLKLILSSFGQAKEMG 614
      . * .

Molgula.oculata.noto6      QDTQHPIQIIIPISGDLTNTVETLNNFKEVE---LEDVCLT-----LI 151
Molgula.occulata.noto6    QDTQHPIQIIIPISGDLTYTMTLNNFKEVK---LEYICLT-----LI 119
Molgula.tectiformis.noto6 QNIQKPVHIIIVPLTGLVNNAKNIMLVFQQTAKL-KEGLCLT-----LV 155
Ciona.intestinalis.noto6  RHVEGRKLIIVPAKITSPIGIVASIDNFVKSQP--KPQVHLV-----LV 240
Lancelet.noto6            RRFWARAALPSASSGFCSTHSHSLTHSLLC-MKXVCLESGAAVSLG 318
Human.noto6              GHNEKVVHILVPLIGRYDIFLRFMENFENMCLIPKQNVKLV-----II 657
      : . : : : * : : :

Molgula.oculata.noto6      SFTGNGMWGYSFDETIIRTHIDKWMLENNNLCAKQQTIVIGDYNVRLAKT- 200
Molgula.occulata.noto6    LFTGNGMWGYSFDETIIRTHIDKWMLENNNLCAKQQTIVIGDYNVRLAKT- 168
Molgula.tectiformis.noto6 LFKKRGYQQQLFDPNIWEYLENFQKNNKEMCFHMREINSDYNFFDGIKQ- 204
Ciona.intestinalis.noto6  LFLGSKNKEISGYKKNLQELTKIKSYQKVNIIQIYPVEGLYSYFKGVRYG 290
Lancelet.noto6            VLFRRSDPEDQTSLDQCKQLDFQYAAKYPAAAIRVEADGEFSRAVGLDT- 367
Human.noto6              LFSRRSDGQD---SSKHIELIKGYKNPKAEMTLIPKMGFESRGLGLEM- 703
      : . : : : . : : . :

Molgula.oculata.noto6      VSSSFDDNSIVLTLKDDIEFGPNVVESCRSAIAPKRKRVFIPLAFAQHDPI 250
Molgula.occulata.noto6    VSSSFDDNSIVLTLKDDMEFSPDVVESCRSAIAPKRKRVFIPLAFAQHDPI 218
Molgula.tectiformis.noto6 SIEEIEENNIILTLKDDIEFNSDAIENCQNLALQNIIRVYFPLSFSQFNPK 254
Ciona.intestinalis.noto6  IKKATISTDVVLLATIEAAFTEQLYHHCQANAIPKKRVYFPIAFGQFNPD 340
Lancelet.noto6            GARQLAPDALLFFCDIDVDVDFQGFQRCRNNAEVRQVYPAVFSQYDPD 417
Human.noto6              ASAQFDNDTLLLFCDVDLIFREDFLQRCRDNTIQGQVYYPPIIFSQYDPK 753
      : : : : * . * : : : * * * . : *

Molgula.oculata.noto6      NIENGMVPGKPKNINKKDINKFTGHWLYQVHDFICAYAADLDSVVQKIFS 300
Molgula.occulata.noto6    NIERGMVPGKPKNINKKDINKFTGHWLYQVHDFVCAYAADFNVLRKTL 268
Molgula.tectiformis.noto6 NIEDGMVPGKPKNVDKKSFTKFTGYWMHNMHDFICAYSDDLKSLSKTES 304
Ciona.intestinalis.noto6  IIKKGMPPGKPKDVNKRDNKFTGYWMHDTDFVCANQADMVGLLNNLSS 390
Lancelet.noto6            LVMEGIP-GKPKPTNLRDINKYNGKDTHKPTNLRDINKYNGKDTHKPTNL 466
Human.noto6              VTNGGNP----PTDDYFIFSKKTGFWRDYGITCIYKSDLLGAGGPDTS 799
      * * : : . * . * . : : :

Molgula.oculata.noto6      EEPEKS-NEIYSEISEYGTAVYN-TFLSKGFKVSSVEPGLLRYYKNHNC 348
Molgula.occulata.noto6    EAPEKS-NEIYFEMSEYGTDMYN-AFLSKGFKVSSVEPGLLRYYKNHNC 316
Molgula.tectiformis.noto6 LVYEENPNNIFIEQSENGDNVYN-KFLANYKVISSVEPGLLRYYKNHNC 353
Ciona.intestinalis.noto6  LGRRMG-----STALGTKIYD-AFLRSDIEVICAVEPGLYRTYQMDPD 434
Lancelet.noto6            RDINKY-----NGKDTHKPTNINKSLTVFRAMPALVHRYHAKTC 506
Human.noto6              IQGWGL-----EDVDLYN-KVILSGLRPFRRSQEVGVVHIFHPVHC 838
      . : : : . : : : :

Molgula.oculata.noto6      SKIKEAPSKRNCIIIRHLESAGSKQALNILYLTEKAS----- 384
Molgula.occulata.noto6    SKIKEATSKRKCIIIRHLESAGSKQALNILYLTEKAS----- 354
Molgula.tectiformis.noto6 TNIRNKLDRKCKIQMMETSQSKQELNILYLNENASKLR----- 392
Ciona.intestinalis.noto6  KGLTRDKSYQHCMRTKAMSLGSKPALGILYINEVLQKKN----- 473
Lancelet.noto6            DPDLTRDQYRMCVAGALSETMASKAQLGLVLLTLQGLKLPAA--- 547
Human.noto6              DPNLDPKQYKMLGSKANTFASTMQLAELWLEKHLGVRYNRTLS 882
      . : * : : * . * : :

```

Figure 3

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Molgula.oculata.noto17 -----
Molgula.occulata.noto17 -----FQSTLSNAKKSFESQNMDPVSASKWSL 27
Molgula.tectiformis.noto17 -----MLLLRCILFVFGFACLNSPIDAKKAKYESQNMDKVSAAEKWDL 42
Ciona.intestinalis.noto17 MLTAYGWIMWKLNLVTLVLLVQVQRTAEDEKPKISLNMDDPVDVKTWGL 50
Oikopleura.dioica.noto17 -----MRILLFIFAAVAALNSSSLKDTTMDKPRLLKFFL 34

Molgula.oculata.noto17 -----
Molgula.occulata.noto17 PDQSSYEIMFRLTDKNYEDVLSESEDPWIVIFY-KSKIDLKWLQHSKIIVG 76
Molgula.tectiformis.noto17 PDKSNYETIFKITDGNYESIMEMEEPWIVLFY-KTKIDRKWLQHSKRVG 91
Ciona.intestinalis.noto17 PLDSKYPDVVQINEANFKPKVLESRDWLVVVF-KDKLHGKWWHHAGHVK 99
Oikopleura.dioica.noto17 PEVDQYIDWVKITDQNFQQRVLESVDPWIVIAPEHQIPVAWKEYATRYR 84

Molgula.oculata.noto17 -----
Molgula.occulata.noto17 GTLWYGKVDLDSNPTLAKKLDNFNDKITNFIGGVGVIFPS-KSSEKMKDLN 125
Molgula.tectiformis.noto17 GAIWYGKVDLEKLNHLANEMNFDHKTTNYVGGIAVVFPSS-KSEKKEIKQFK 140
Ciona.intestinalis.noto17 GAVWYGSVDIEKQENFAKLLAPESTAENIGNGVAVIFPFVTPKEKIKFT 149
Oikopleura.dioica.noto17 GQFWFGKFFNKWGENLEERFGEYEGKPK-----VLVYSWTREAKKKGPFI 128

Molgula.oculata.noto17 -----AENMAAETLPDWTTKFDFTPATMPKFNNEWVINSYY 35
Molgula.occulata.noto17 SKKLLK---VIEPLAENMAAETLPDWTTKFDFTPATMPKFNNEWVINSYY 172
Molgula.tectiformis.noto17 AKKLIK---VSEPHLAEKFASETMEDYTTELDFLPSNMQVFNDDVWVNTYY 187
Ciona.intestinalis.noto17 GKKKRRLIVKSPTKAEEIISSIPNQIERMNFKPSRMKFFQDQVWVDSYY 199
Oikopleura.dioica.noto17 TDNFSV---ALEKAIESLPVTSLQRLS--FDTTIEDPSNIDAFLTNALY 172
      *.:  ::  ::  ::  ::  ::  ::  *

Molgula.oculata.noto17 QSEI-TRYPVICVS--DEPG--VPLLKSLSVHFKSHFSFAVVSASVPR 80
Molgula.occulata.noto17 QSEI-ARYPVMCVT--DEPG--VPLLKSLAIHFKSHFTFAVVSASVPR 217
Molgula.tectiformis.noto17 KSPI-AKFPILCVT--DTGD--VPAVVKSLSKLMSHFVIGVQKSSAHR 232
Ciona.intestinalis.noto17 KSSP-PRFVLCIT--DEPEPTIPVSMVLVLSNYFSRFFSFALVRKEDVSR 246
Oikopleura.dioica.noto17 GEEKNPKWPVIFLLGKEDDEEGLPPIITRIVISHYFSDSFKFAFVKFDQLKA 222
      .  .:.*::  :  :  :*  ::  :.  *  .:.  *  .

Molgula.oculata.noto17 MIANFKEVP---ESTQFPMFYLLMGREPSSDEMNRGLYMNMFNIFPYIKE 127
Molgula.occulata.noto17 MMANFKEVP---EPTHPMFYLLIGREPSTDEMNRGMYNMFNIFPYIKE 264
Molgula.tectiformis.noto17 MISIFKEVP---MPKVFPMYTLLGREPTSEEMNRGSYKLFHDFIFPYLSE 279
Ciona.intestinalis.noto17 MRLQFKEVP---EPKVYPHYLMLMGKESPEEMDRGSGFMHFNIFPFLAD 293
Oikopleura.dioica.noto17 LKKVFPDYPTDMKDEFPTFIVAGKEPS-DDAPEDEFELTFVVRMSKQ 271
      :  *  :  :*  :  :*:  ::  ..  :  :  *  :.  :

Molgula.oculata.noto17 KYGDSTIFHNVANYLFFVANHDFRSMPLGKRSDE----- 160
Molgula.occulata.noto17 KYGDSNIFHNVANYLFFVANHDYRSMPLGKRSDETEKVKMKMGI SQALSQR 314
Molgula.tectiformis.noto17 KYGDSSLFHNVLNYMFLVNQDYRADLPGKRRGESSAIKNMEEIKEKMNRR 329
Ciona.intestinalis.noto17 KYGRSELFVSVKFLFTANEDYRNEPGRRSDEAISEARMSLVDRSLRNR 343
Oikopleura.dioica.noto17 KFGMSTKFNDAVAFFFWVNEQIRETLPGRDPNEDMDDKKTMGSI RNKLWKR 321
      *:*  *  *  *  ::*  .*:  *  ***:  .*

Molgula.oculata.noto17 -----
Molgula.occulata.noto17 VEILMK----- 320
Molgula.tectiformis.noto17 IKLLLKSQHKKLNKSEL----- 346
Ciona.intestinalis.noto17 VELVNHKFRKQKNVEL----- 359
Oikopleura.dioica.noto17 LEI IKPAVMKAQKESDVMQAARKFENLNAASKLLEEKEKEAERLKNELE 371

Molgula.oculata.noto17 -----
Molgula.occulata.noto17 -----
Molgula.tectiformis.noto17 -----
Ciona.intestinalis.noto17 -----
Oikopleura.dioica.noto17 ETQAKLEEAEKGKIAESEDNSTEDPADAKADVEQKDEL 409

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Figure 4

```

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  MIQNKKKMLIESVICCILSIIIAANRIQATSVTYQPFEENFETGVYAYWN 50
Ciona.intestinalis.leprecan -----
Human.leprecan -----

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  NDWENAIVEIEKSLVMHRLFESGIYDCYMKCKDTRNIENTDRHSQFADIK 100
Ciona.intestinalis.leprecan -----
Human.leprecan -----

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  FIISKASCSSRCLDEIFGDARLITQVSEHIKHLFHIKRPYVYLQFAYFQV 150
Ciona.intestinalis.leprecan -----
Human.leprecan -----

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  GNIEQAVKTAHTYWLHRVGGDDVIEDSIEYYRNLTVEVKPEFFIDLEASEHI 200
Ciona.intestinalis.leprecan -----
Human.leprecan  -----MEMQQNIENYRATAGVEALQLVDREAKPHM 30

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  YVYDTAVEIYLDDEYEAAILPFMRALEAYYTASDKCNALCEG-EHVFDDYD 249
Ciona.intestinalis.leprecan -----
Human.leprecan  ESYNAGVKHYEADDFEMAIRHFEQALREYFVEDTECRTLCEGPFQRFEEYE 80

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  YP-DVPEFHMQTADHYIQVTECSLECVKKIAT-DSSGVHISDFLPLHYHY 297
Ciona.intestinalis.leprecan -----
Human.leprecan  YLGYKAGLYEAIADHYMQVLVCQHECVRELATRPGRLSPIENFLPLHYDY 130

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  LQFAIYKEGNI SAALSNAKTYLLFHPPEDEVMVNNVNLSSKKVDSKTEAIP 347
Ciona.intestinalis.leprecan -----
Human.leprecan  -NVADSTENNADLAEN-----EDSIVGEMGDISKNTEDEDNGD 52
LQFAYYRVGEYVKALECAKAYLLCHPDDDEDVLDNVYDYESLLDDS----- 175

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  EAVLYKKRIDGQIALRLYSYQAFGYEYRTNLLTPFADLTENEDGIDFFS 397
Ciona.intestinalis.leprecan -----
Human.leprecan  SAENIELKEN-----SDKSTKNVAA 72

Molgulid.hybrid.67122.leprecan -----
Molgulid.hybrid.67123.leprecan -----
Molgula.tectiformis.leprecan  -----EPETEELAPEDSFYQSFQEKQADIIEKKS IENEILKEIEAKKTES 45
Ciona.intestinalis.leprecan -----
Human.leprecan  -----EPETEELAPEDSFYQSFQEKQADIIEKKS IENEILKEIEAKKTES 45
GHTLQKPNILPSNPEEQFYQSPDEKQKDIIEKKSMEEHILNELEEQRSQ 447
TLKQVEPPKESKTQAKVEYNSMKDKTEDMIEAKDAYNARMEVIRKKRE-- 120
----IDPASIEAREDLTMFVKRHKLESELIKS-AAEGLGFSYTEPNYWIR 220
      . *           : . . .   :*:      : . . :

Molgulid.hybrid.67122.leprecan  TTRETEDDV-VDDARSNTHLITSEG MVTIGRTDSVS--SNAEGKILSEKT 92
Molgulid.hybrid.67123.leprecan  TTRETEDDV-VDDARSNTHLITSEG MVTIGRTDSVS--SNAEGKILSEKT 92
Molgula.tectiformis.leprecan  KVAKPDNDVDVSNVSLGEALVDSKDMVTIRDAANET--PTAEGNLLFEDY 495
Ciona.intestinalis.leprecan  --AESDVDISHMSSDIGHVEIDSNSIIISIIDSTSLPQPPTPEGPLLNKV 168
Human.leprecan  YGGRQDENRVPVSGVNVGEAEVHGFSMGKCLKSPKIDR-DLREGGPLLYENI 269

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Human.merlin QMKQAQAREEKARKQMERQRLAREKQMREEAERTRDELERLLQMKEEAT 368
 **** **:***::** *: ::

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin MAMEALNRSEETAHELLAEKAQIAEEETVLLKKKANHAEQEMQRIKAEFAK 432
 Ciona.intestinalis.merlin SAADTLRRSEETAELLGEEKAKVAEEEAQLLRAKLNKSDQETIQLKLEIGK 439
 Acorn.worm.merlin YANEALMRSEETADLLGEEKASVAEEEAALLAQKATEAEQETQRIKMQAIK 408
 Human.merlin MANEALMRSEETADLLAEKAQITEEEAKLLAQKAAEAEQEMQRIKATAIR 418

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin TQEANIQLQONLQNYDQVTKQLAEDSKTNAEAEKLRKELKRAMQAKNEA 482
 Ciona.intestinalis.merlin LQEANHLQPKQFLKYEQYVSQLEQAQARAGELKNVREELYATKAALNDA 489
 Acorn.worm.merlin SEEEKMLMEQKALEAEMIASKIAKDSERRNKEADQLKEELIRAKEAEKSA 458
 Human.merlin TEEKRLMEQKVLAEVLAALKMAEESERRAKEADQLKQDLQEAAREAERRA 468

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin KEKLTNASMVNSNNTSTDMSYGHYIPDTPGMASPLSNAMGESVMVTGAS 532
 Ciona.intestinalis.merlin NIKLN---LLANQHVHSPNTSNSVPMN--MGFLPINNQIHSQVISMGP- 533
 Acorn.worm.merlin KNRLIEITRTWTTP----- 474
 Human.merlin KQKLEIA--TKPTY----- 482

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin VSPLQIQPLAPGVPIVSHKHIGSNGSKASNGSIGGANNGGIGQNGRPGNV 582
 Ciona.intestinalis.merlin -INLNNTSQSNHSPAVSN-AQYYHMYAASNGSAHSVKTGDSNVTDNRNSNE 581
 Acorn.worm.merlin -----MYDVMSHDLSDLHLDLDTANSEFACDLMTHG----- 506
 Human.merlin -----PMNPIPAPLPDIPSFNLIIGDS-LSFDFKDT----- 512

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin GMHKVGSNGGLQTIIGDYDDSDNGDFARVSEMQLSQEIEKERMEYQAKSK 632
 Ciona.intestinalis.merlin -----FTSAPGN----SDMQQLSQEIEKERMEYHVKSR 610
 Acorn.worm.merlin -----DVEQLSLEIEKERVEYMEKSK 527
 Human.merlin -----DMKRLSMEIEKEKVEYMEKSK 533

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin HIEQQLAMLKTEIEGLKVDEKMTPLDHMYTENS�KG-AVKYQSLNQAKES 681
 Ciona.intestinalis.merlin NIEQQLFNLRSIEIEVLKVDSEMTGFDQK--QDSNQP-HTHEISTFQGHKE 657
 Acorn.worm.merlin HLQAQLNDLKTEIEVLKIEEKQTHLDQLHNSMVQKG-DTKFSTLKKIKAG 576
 Human.merlin HLQEQNLNELKTEIEALKKERETALDILHNENSDRGGSSKHNTIKKLPQL 583

Molgula.occulta.merlin -----
 Molgula.oculata.merlin -----
 Molgula.tectiformis.merlin TPEQRLQLYNKL 693
 Ciona.intestinalis.merlin TPQYYDGL---- 665
 Acorn.worm.merlin TTRARVAFFEEEL 588
 Human.merlin SAKSRVAFFEEEL 595

Figure 6

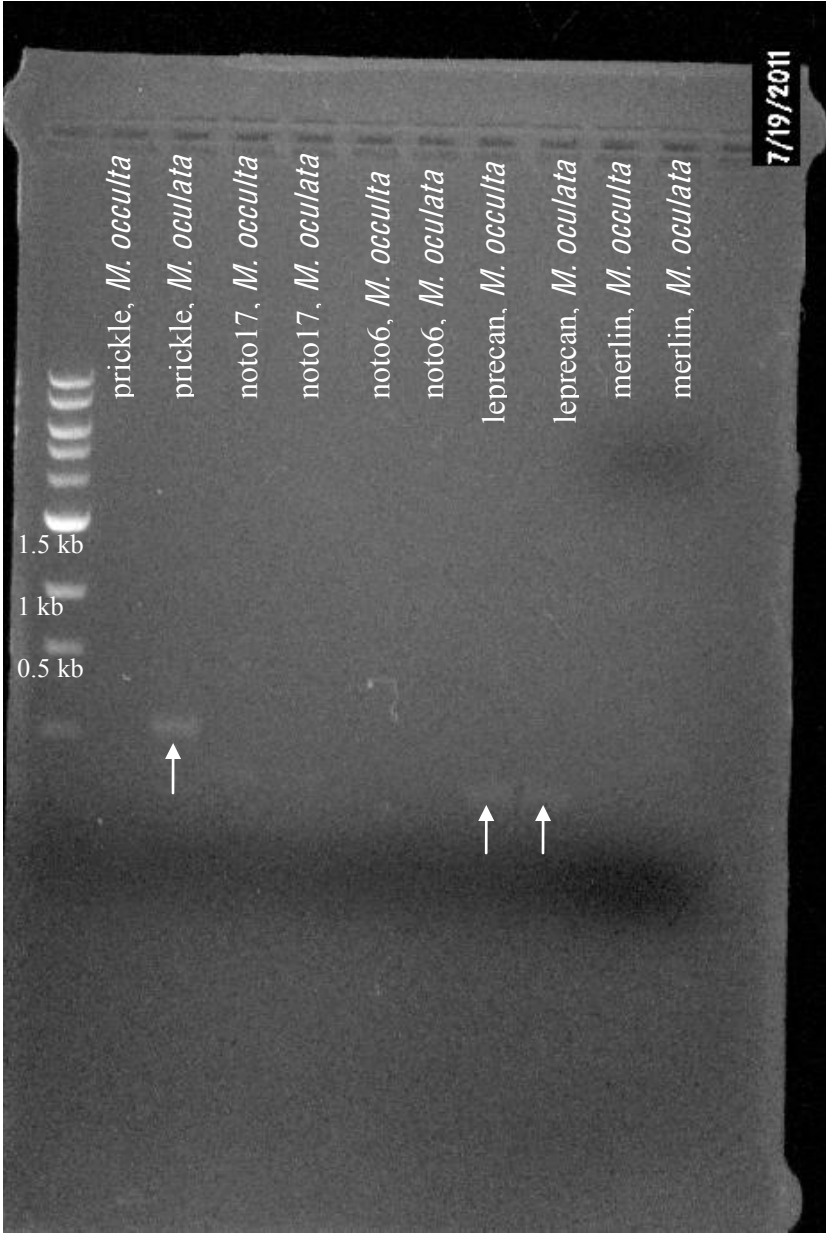


Figure 7

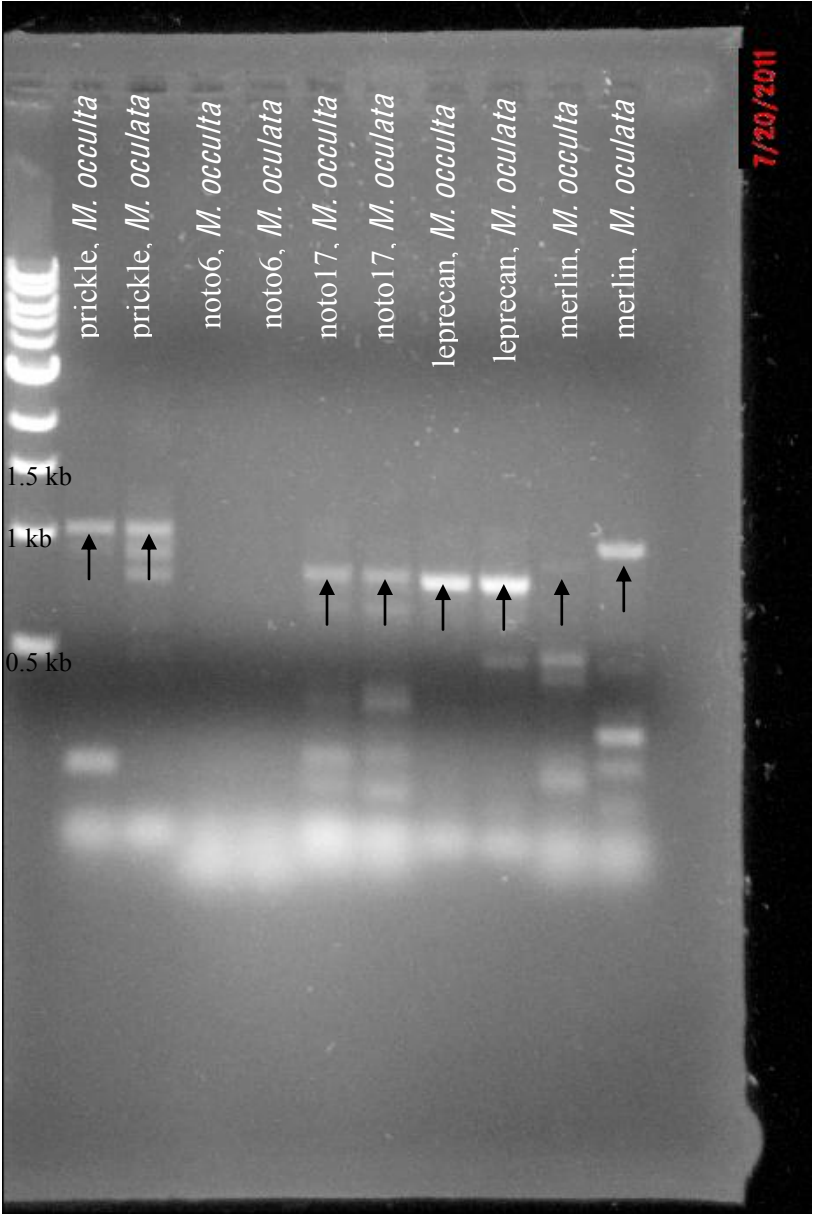


Figure 8

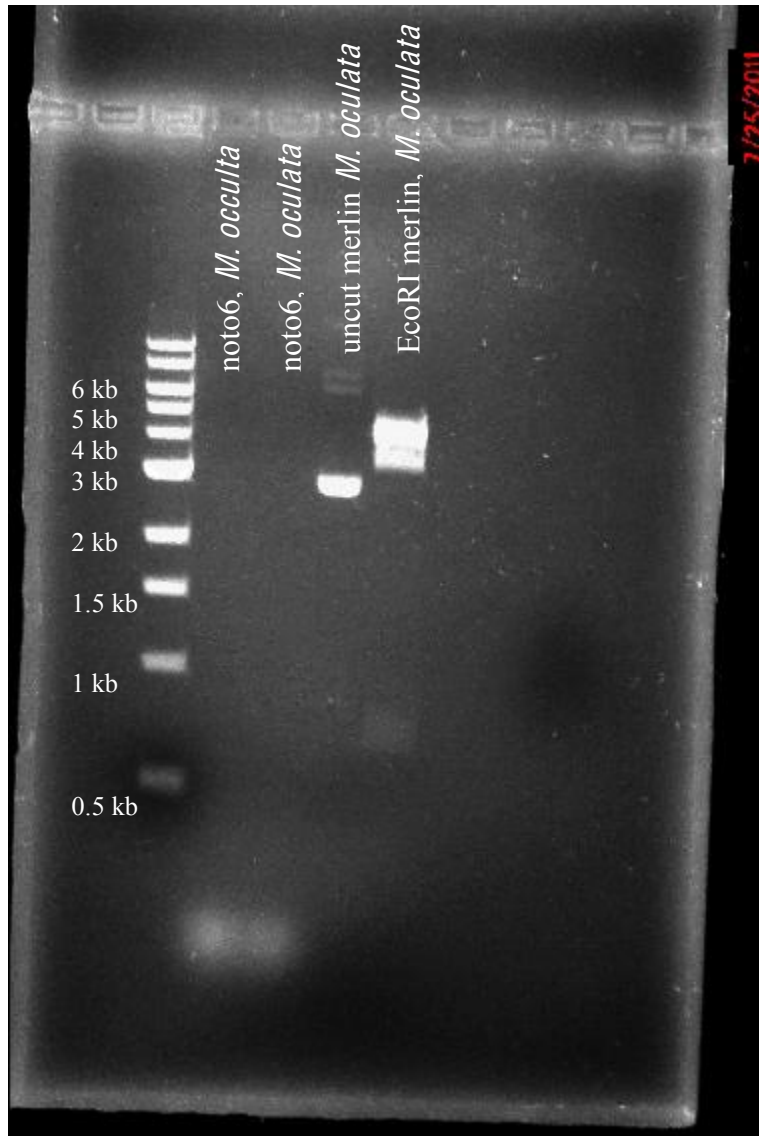


Figure 9

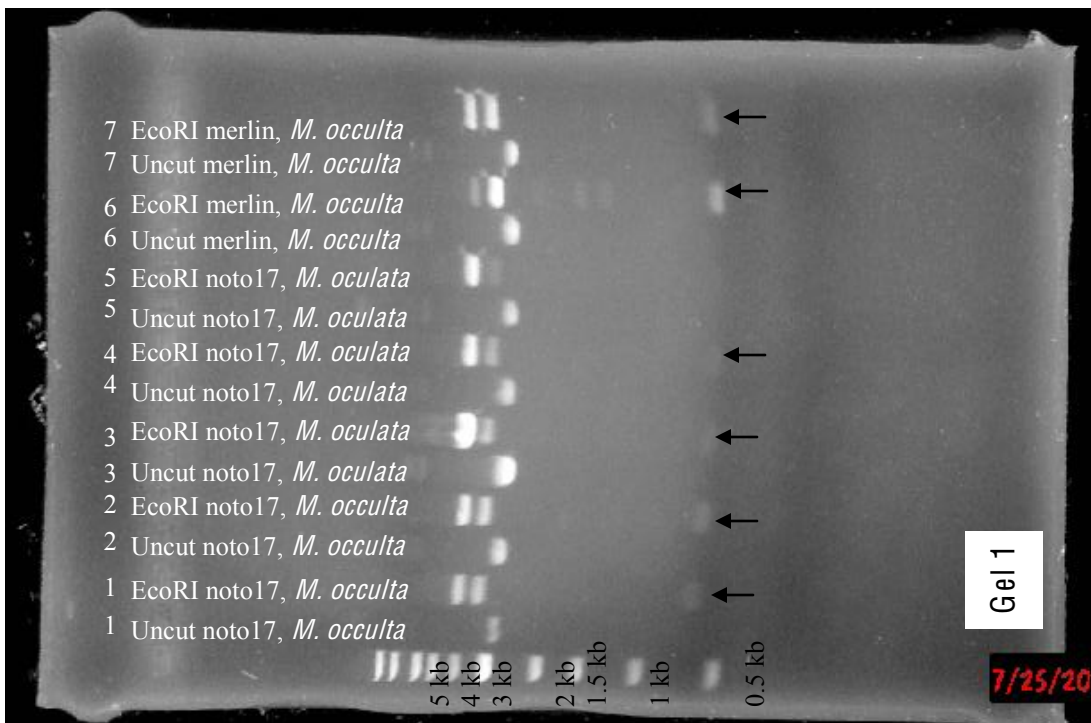
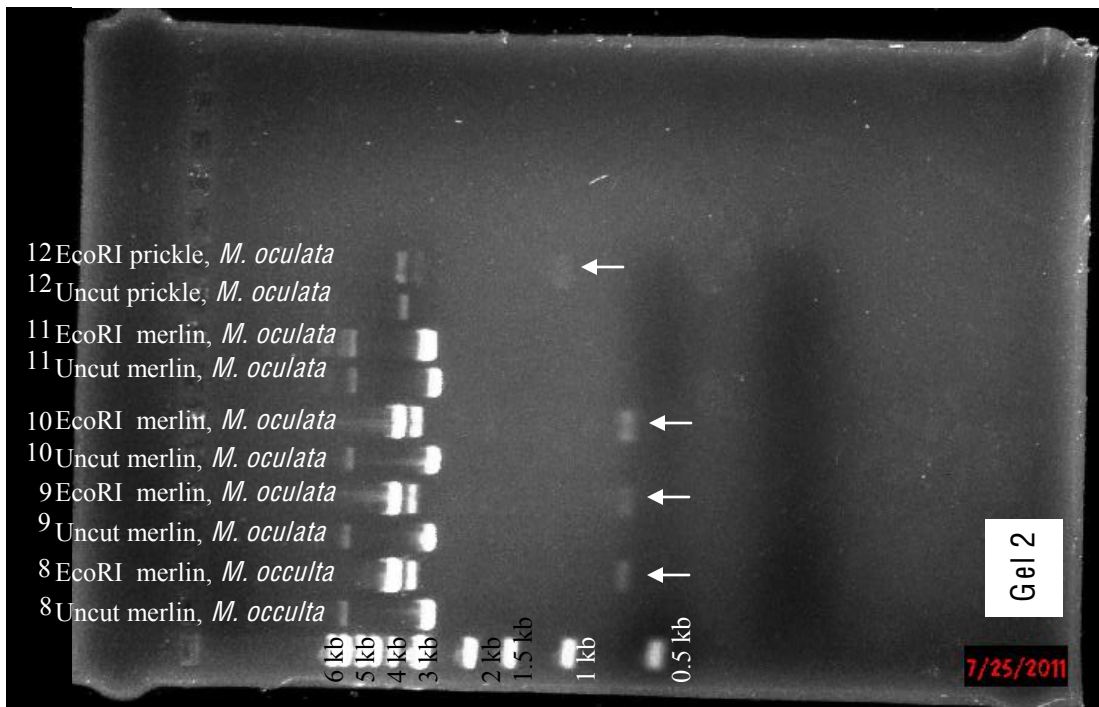


Figure 10

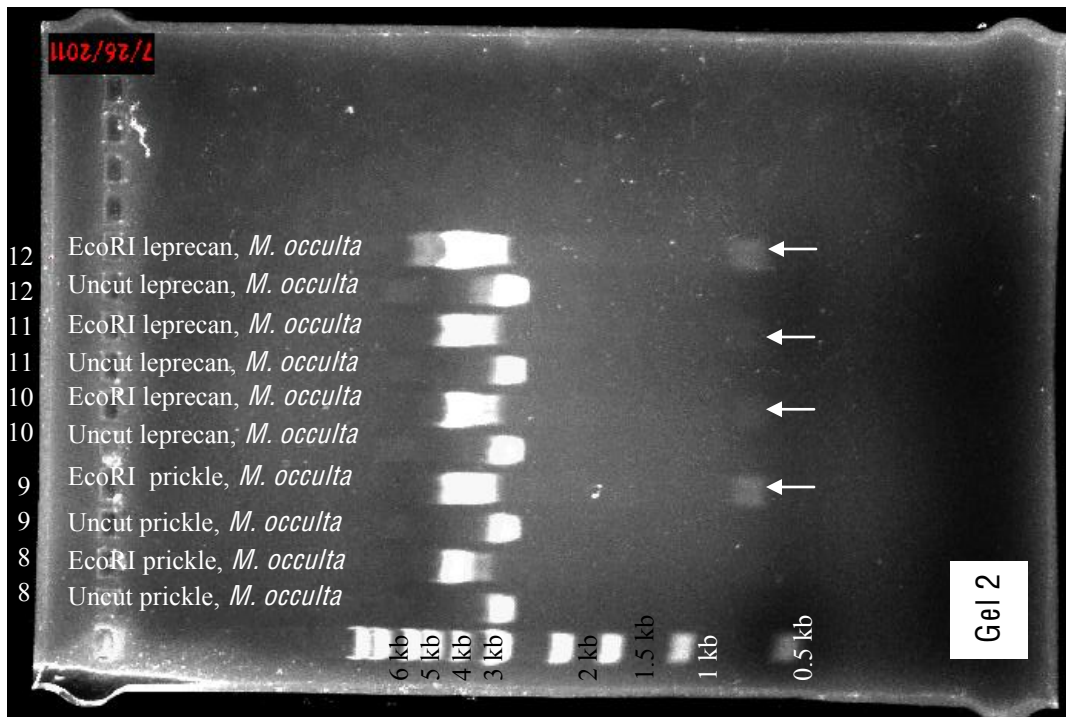


Figure 11

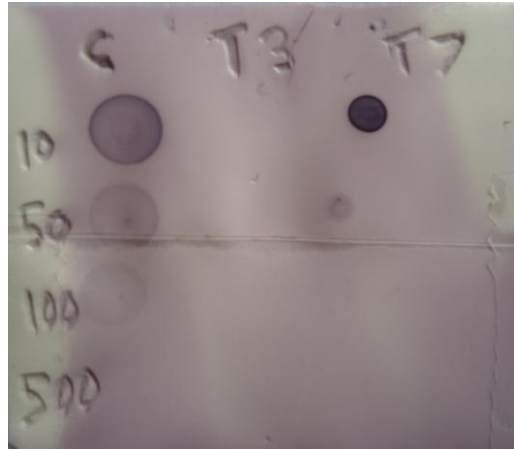


Figure 12

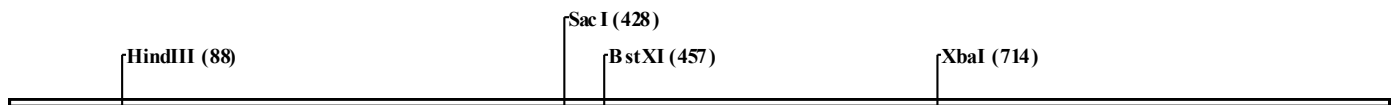


Mapping all cutsites.

Cutters : BamHI, HindIII, KpnI, PstI & XbaI

Non-Cutters : ApaI, Bsp106, BstXI, DnaI, EcoRI, EcoRV, NotI, SacI, SacII, Sall, SmaI, SpeI, XhoI & XmaIII

Figure 13



Mapping all cutsites.

Cutters : BstXI, HindIII, SacI & XbaI

Non-Cutters : ApaI, BamHI, Bsp106, DnaI, EcoRI, EcoRV, KpnI, NotI, PstI, SacII, Sall, SmaI, SpeI, XhoI & XmaIII